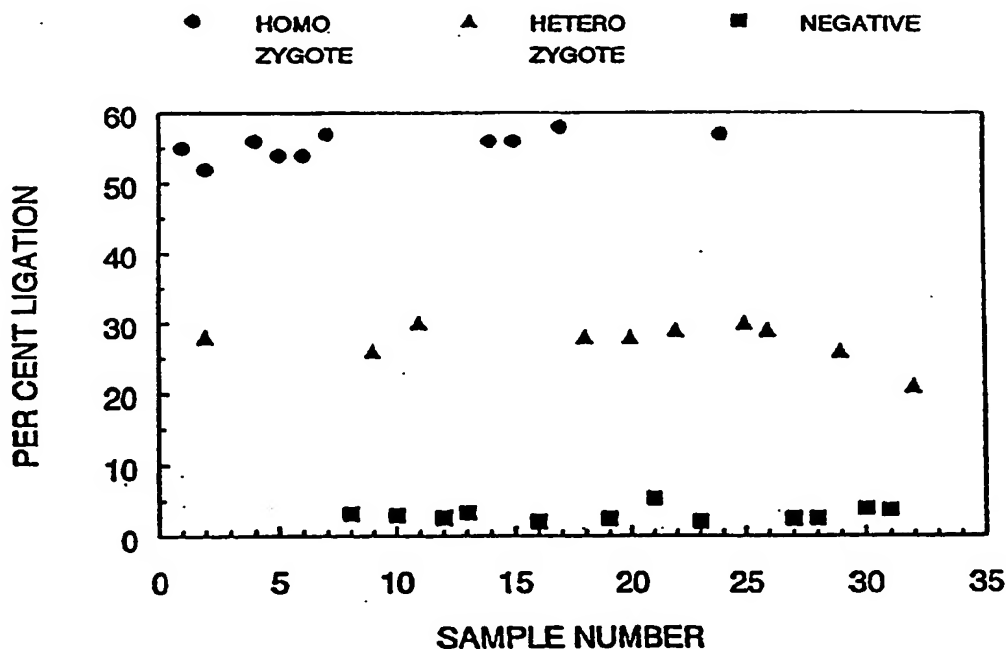




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(54) Title: HIBRIDIZATION-LIGATION ASSAYS FOR THE DETECTION OF SPECIFIC NUCLEIC ACID SEQUENCES



(57) Abstract

A new method has been developed for conducting a gene probe assay. The preferred technique involves (1) using a gene amplification technique (e.g., PCR) to multiply the gene sequence of interest and (2) using a hybridization-ligation detection methodology, wherein the sequences of probes hybridized to the target sequence allow for separation and detection (e.g., probes might contain a combination of magnetic particles and acridinium esters) to determine if a specific sequence is present.

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HYBRIDIZATION - LIGATION ASSAYS FOR THE DETECTION OF SPECIFIC NUCLEIC ACID SEQUENCES

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Background of the Invention

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Gene probe techniques have become an important analytical tool for predicting the incidence of inherited disease and in diagnosing existing medical conditions. However, the currently-used techniques are slow, laborious and involve the use of harmful chemicals. Some procedures used in the gene probe field were discussed recently by Plaha et al (14 BioTechniques 566, 1993). Current gene probe techniques typically involve the use of electrophoresis, frequently on long polyacrylamide gels. Several of the chemicals to which the lab personnel are exposed are considered to be hazardous. Namely, the acrylamide monomer, some of which may remain in the polymeric gel, is considered to be a neurotoxin. The ethidium bromide used as the staining agent is a mutagen. Polyacrylamide gel electrophoresis typically requires several hours, while the analysis using the new gels described in Plaha require 16 hours for a single run. Other techniques use radioactive markers, which require the use of special handling controls and disposal techniques. Also, the other aspects of the current procedures require a relatively long period of time. (For

example, the Southern Blot procedure requires about 48 hours to complete.)

5 Furthermore, the electrophoresis technique itself may
produce uncertain results. First, ethidium bromide
techniques are often not very sensitive. Second, the
conditions to adequately separate the different gene
fragments must be developed. Third, the results are
nearly always qualitative, not quantitative. In addition,
10 the inability to distinguish fragments of DNA which are
the same or similar size but differ in sequence, even by
a single base, also limits the usefulness of past
procedures.

15 Limitations on the sensitivity inherent to DNA based
assays may be overcome by the polymerase chain reaction
(PCR). The amplification of a specific sequence by PCR
enables the detection of that sequence when present in a
sample in extremely low amounts (Saiki, et al., 230
20 Science 1350, 1985). Although the PCR technique can
amplify DNA sequences to overcome the sensitivity
limitations that existed before PCR was available, a
number of problems remain attendant with the use of PCR.
The products of a PCR reaction often include artifacts due
25 to primer-dimers and non-specific priming events,
especially in the absence of the target sequence in the
sample. By virtue of its exquisite sensitivity, PCR is
susceptible to false positive results due to
contamination. The PCR technique itself does not readily
30 allow the discrimination of small differences between
sequences such as point mutations which may underly
genetic diseases, such as in cystic fibrosis. As a
consequence of these limitations, verification of the
presence of the specific target sequence after

amplification by PCR is a desirable if not an essential step in a DNA assay.

5 Earlier workers have used hybridization and ligation techniques as a precursor to analyzing samples on various gels. (See, Landegren et al, 241 Science 1077, 1988). However, these techniques are slow, inconvenient and are not amenable to usage on automated instruments.

10 A novel analytical method has been developed, which eliminates the drawbacks of the current techniques for the analysis of DNA sequences and provides quicker and more accurate results. The instant novel technique can be utilized along with PCR to improve accuracy in gene probe
15 assays.

Summary of Invention

20 A new method has been developed for conducting a gene probe assay. The preferred technique involves (1) using a gene amplification technique (e.g., PCR) to multiply the gene sequence of interest and (2) using a hybridization - ligation detection methodology, wherein the sequences of
25 probes hybridized to the target sequence allow for separation and detection (e.g., probes might contain a combination of magnetic particles and acridinium esters) to determine if a specific sequence is present.

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Brief Description of the Drawings

Figure 1: The probe assay formats used for the detection of the delta F-508 and normal alleles.

Figure 2: The results of the HLM for the simultaneous detection of delta F-508 and normal alleles in nine samples of human DNA that have been amplified by PCR. The probe specific for the delta F-508 allele was labeled with DMAE and the probe specific for the normal allele was labeled with LEAE.

Figure 3: A portion of the sequence of exon 10 of the CFTR gene in the vicinity of the sites for the delta F-508 and delta I-507 mutations. Sequences underlined are complementary to the sequences of probes immobilized on paramagnetic particles (PMP.508 or PMP.507) and labeled with acridinium ester (508.NOR or 507.NOR).

Figure 4: The hybrids formed between the probes for the delta F-508 assay and delta I-507 assay and the various alleles.

Figure 5: HLM results for the detection of the delta I-507 mutation in nine samples of human DNA that have been amplified by PCR.

Figure 6: HLM analysis of three samples of human DNA amplified by PCR for the presence of the normal, delta F-508, and delta I-507 alleles.

Figure 7: HLM analysis of three samples of human DNA amplified by PCR using the simultaneous hybridization-ligation protocol with Taq DNA ligase for the presence of the normal, delta F-508, and delta I-507 alleles.

Figure 8: HLM discrimination of the G542X and Normal alleles.

Figure 9: Simultaneous detection of delta F-508 and G542X mutations by HLM.

5 Figure 10: Discrimination of normal, G551D, G551S, and Q552X sequences by HLM with ^{32}P -G551D.NOR or ^{32}P -G551D.CF using T4 DNA ligase with 200 mM NaCl.

10 Figure 11: HLM with ^{32}P -G551D.NOR and ^{32}P -G551D.CF using T4 DNA ligase with 600 mM NaCl.

Figure 12: Discrimination of normal, G551D, G551S, and Q552X sequences by HLM with ^{32}P -G551D.CF or ^{32}P -G551D.NOR using Taq DNA ligase with 200 mM KCl.

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Figure 13: Comparison of assays for G551D with Taq DNA ligase under different salt conditions.

20 Figure 14: The p53 model for systematic evaluation of ligation specificity. The top sequence is that of the target (region flanking codon 175 of the p53 gene). The probe sequences are on the bottom. The positions marked "X" and "Y" were systematically varied with the four nucleotides.

25

Figure 15: Calculated per cent ligation for the delta F-508 assay of PCR amplified samples.

30 Figure 16: The correlation of per cent ligation calculated from HLM with the genotype of PCR amplified human DNA. The error bars represent the 99% confidence interval.

Figure 17: Discrimination of the delta F-508 and delta I-

507 sequences with delta F-508 probes by T4 DNA ligase as a function of NaCl concentration.

5 Figure 18: Figure 18 represents the discrimination of 5' (2) and 3' (2) mismatches using T4 DNA Ligase and Taq DNA Ligase, in terms of per cent ligation.

10 Figure 19: Figure 19 shows the 3' (1) mismatch discrimination for different base pairs, in terms of percent ligation. Data is shown for 2 different concentrations of T4 DNA Ligase (1 nM and 240 nM).

15 Figure 20: Figure 20 shows the 5' (1) mismatch discrimination for different base pairs, in terms of percent ligation. Data is shown for 2 different concentrations of T4 DNA Ligase (1 nM and 240 nM).

Detailed Description of the Invention

20 The novel technique has been developed for gene probe analyses. This first step of DNA analysis generally involves using an amplification (e.g., PCR) technique to multiply the sequence of interest. Obviously, if sufficient quantity of the unknown sequence is present in
25 the assay sample, amplification may not be needed. Following the amplification process, a hybridization-ligation methodology (HLM) is used to confirm the identity of the amplification product. To help identify the gene sequence, an easily separable
30 particle (for example, a magnetic particle) is used along with an identifiable moiety (for example, a luminescent marker such as an acridinium ester).

The initial part of the technique involves the use of

an amplification procedure to multiply the sequence being investigated if insufficient quantity of the sequence to be identified is present. PCR techniques have been known for several years. For example, Saiki et al described an enzymatic amplification technique for β -globulin genomic sequences by providing 2 oligonucleotide primers flanking the region to be amplified, annealing the primers to strands of denatured genomic DNA and extending them with a DNA polymerase from *E. coli* or *Thermus aquaticus* and deoxyribonucleosidetriphosphates, and repeating cycles of denaturation, annealing and extension (Saiki et al (230 Science 1350, 1985); Saiki et al (239 Science 487, 1988)). Various amplification techniques have been developed recently (e.g., ligase chain reaction (LCR) and Q β Replicase), and any of the amplification techniques can be used instead of PCR or in combination with one or more other amplification techniques. Furthermore, it is anticipated that other amplifying techniques will be developed. The exact technique used for amplification is immaterial to the invention herein, and those with knowledge in gene probe procedures are assumed to be familiar with the overall techniques used therein and the reasons for preferring one technique over another. The critical fact is that the relatively small sample of DNA is amplified so that the analytical technique used thereon is more sensitive than it normally would be.

Typically PCR techniques involve the procedure described above. There are many variations of this basic technique known to those with skill in the art, for example those described in PCR Protocols (eds. Innis, MA, Gelfand, DH, Sninsky, JJ, and White, TJ, Academic Press, 1990) . Details of one example of how a PCR reaction is run is described in detail in the examples herein.

Once an amplified sample is available, this material is analyzed by HLM. In HLM, the sequence which is complementary to some or all of the target sequence is incorporated into 2 or more probes which are reacted with the target oligonucleotide. Much of the discussion here relates to the use of only 2 probes, but more than 2 can also be used, as discussed below. One portion of the complementary sequence is attached to an insoluble material that can easily be separated from a reaction mixture. For example, a magnetic particle might be used. Another possible material is a material that can be separated by centrifugation from the reaction mixture. The second portion of the complementary sequence is attached to a material that can be detected by an analytical technique. For example, a chemiluminescent material, such as an acridinium ester, might be used. Other examples are fluorophores or chromophores. These 2 complementary sequences are hybridized to the target sequence. The hybridization solution contains salt, typically approximately 500 to 700 mM NaCl, with concentrations of about 600 mM being most preferred. The hybridization is carried out at elevated temperature (e.g., 45°C.). After hybridization, the insoluble properties of one of the probes is used to separate the hybridized from unhybridized probe with the label. A ligase is then used to attempt to join the 2 complementary sequences of the probes. If the 2 complementary sequences precisely match the target in the immediate region of the junction of the two probes, the terminal nucleic acids are close enough to each other to be connected by the ligase. On the other hand, if the target sequence is sufficiently different from the suspected target, the terminal nucleic acids on the probes are sufficiently far from each other so that they cannot be joined by the ligase. For example,

if the target has a deleted nucleotide at the place where the 2 probes meet, the terminal nucleotide on one probe will overlap the other probe, and the two probes will not be ligated. Similarly, if the target has an inserted nucleotide, the 2 probes will not be close enough to each other to permit ligation to occur. Furthermore, if there is a mismatch at the location where the 2 probes meet, the 2 probes will not be ligated efficiently under conditions defined herein.

Although the ligation proceeds as described above, depending on whether the terminal bases of the probes couple or fail to couple with the target, those with skill in the art will recognize that the mismatching of bases on the probe away from the terminal positions may also have some effect on the binding of the probe to the target. For example, if a base located on the probe several base positions away from the terminal base fails to bind with the corresponding base on the target, there may not be sufficient discordance between the probe and target to prevent the terminal probe bases from being ligated. Steric factor and the totality of binding between the probe and target will have an effect on whether the instant technique is totally effective in determining the composition of the target. On the other hand, it should be noted that discordance away from the junction of the probes may have a sufficient effect so that the two probes will not be ligated. An example below shows a case in which discordance at a site 2 bases away from the junction of the probes causes sufficient interference that the probes were not ligated.

Similarly, by using sets of probes that are expected to hybridize and ligate to a portion of the normal

sequence, it is possible to determine whether the target contains a mutation. If it is found that hybridization or ligation do not occur, it can be concluded that the mutation probably occurs in that portion of the target being examined. By moving to the next portion of the target, a similar experiment can be run. Thus, by moving sequentially along the target, one can determine the site or sites on the target where mutations are found and then proceed to design experiments to identify the exact mutation which occurs at each of the mutation sites.

The ligation is carried out under conditions which will ensure the specificity of the reaction (see examples below). Ligation can be carried out using one of the many ligating reagents available, such ligating reagents typically achieving ligation by chemical or enzymatic action. One important difference between the conditions utilized here vs. the prior art is that a much higher salt concentration has been found to assure ligation specificity. The previously used salt concentration (200 mM NaCl) had been found to permit ligation of mismatched probes. In the instant invention, it has been found that higher salt concentrations yield unexpectedly improved specificity. For example, to attain the most specific ligation with T4 DNA ligase, the salt concentration is typically approximately 500 to 700 mM NaCl, with concentrations of about 600 mM being most preferred and concentrations up to about 1000 mM being usable.

Variations in the ligation process are possible. For example, many different ligating agents can be used, and examples showing the use of T4 DNA-ligase and Taq DNA-ligase are shown herein. Furthermore, it may be preferred to include in the Taq ligase buffer other components which

may increase the sensitivity of the reaction. For example, it has been found that the inclusion of tRNA reduces background signal which is caused by non-specific bonding of the labeled probe.

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After the ligation step is undertaken, a denaturation step separates the target sequence from that of the probes and ligated from unligated probe. The material connected to the insoluble material can then be separated from the reaction mixture by centrifugation, application of a magnetic field or other appropriate procedure, and the presence of any label connected to the insoluble material due to the action of the ligase can be determined.

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By using the disclosed components, the separation of the target material can be achieved using a technique other than chromatography or electrophoresis. Thus the technique can be accomplished much faster than if electrophoresis or chromatography were used. Furthermore, the detection technique can be a more quantitative one, such as the measurement of radioactivity, fluorescence or luminescence. Other detection methods can utilize commonly available techniques that permit the subsequent addition of the label. For example, the probe might have chemically attached thereto biotin, and, after separation of the probes, the label, being bonded to avidin or streptavidin, can be reacted thereto, thus forming a probe linked to a detectable label. Thirdly, the technique can now be utilized in some of the automated instruments, such as the ACS 180 instrument manufactured by Ciba Corning Diagnostics Corp. of Medfield, MA..

It should be noted that a variation of the instant technique can be used to determine information about the

sequence on the target. If, after denaturation, it is found that no label is connected to the insoluble probe, a different aliquot of the reaction mixture which has not been denatured can be further analyzed. In that sample, 5 the insoluble marker can be separated (e.g., by the application of a magnetic field, by use of centrifugation, etc.), and the target can be analyzed to determine if the marker probe is attached to the target. If this is found to be the case, the sequence of much of the target 10 polynucleotide can be predicted due to the hybridization of the 2 probes to the target, and further experiments to confirm the sequence in that region of the target (e.g., near the ligation point) can be planned.

15 Alternatively, a similar analysis can be conducted by denaturing the sample after ligation and then separating the solid phase. In this technique, both the separated solid phase and the supernatant are analyzed for the presence of the labeled probe. If most of the label is 20 found only in the supernatant, it can be concluded that ligation of the probes did not occur, which is an indication of a mismatch at the expected ligation point. However, since the labeled probe became attached to the target, it can be concluded that the target had the 25 expected sequence, or a sequence close to expected, or else the labeled probe would not have hybridized to the target. Thus, even though ligation has not occurred, much can be inferred about the sequence of the target. Furthermore, the sum of the amounts of the label in the 30 supernatant and on the solid phase should approximate the total amount of target in the assay sample. The percentage of the labeled probe which is ligated should indicate the homozygosity or heterozygosity of the sample. In addition the ratio of label on the solid phase to the label in

5 solution can indicate more information about the sequence on the probe, for example the existence of diseases wherein portions of genetic material are replicated (e.g., fragile X). It should be noted that, in conducting these assays, the data have not been found to be exactly the theoretically expected values (i.e., not 100% of the label is found in the solid phase for a homozygous sample). (See example below.)

10 Thus, by knowing the potential mutations that can occur at a particular site, it is possible to generate a specific probe so that the sequence on the target can be confirmed. If several potential mutations can occur at one site, it is also possible to design several probes,
15 each with a different label, to determine whether it is the normal sequence, and, if it is a mutated sequence, which of the mutations occurs. Similarly, mutations that occur near each other in a target sequence can be determined. In addition, the technique of using two or
20 more differently labeled probes can be utilized in the case where multiple forms of the target are expected, such as in the fragile X case discussed above.

25 The two different labels used in the same assay may be, for example, a fluorescent donor and fluorescent acceptor pair. In this case, by varying the incident light, the two labels can be used to distinguish among three possible outcomes. If the incident light to the first probe gives fluorescence typical of the first label,
30 this is an indication that only the first target is present. If the fluorescent output is that from the second label, two alternatives are present. If incident light which excites the first label gives fluorescence from the second label, this is an indication that both

targets are present. On the other hand, if only the incident light which excites the second label gives fluorescence from the second label, this is an indication that only target two is present.

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For example, multiple mutations in one vicinity have been found in variations of cystic fibrosis. For example, the delta F-508 and delta I-507 mutations are both 3 base pair deletions, the positions of these mutations are partially overlapping in the sequence of the CFTR gene. (See Zielenski et al (10 Genomics 214, 1991) for sequence of the CFTR gene.) Analysis of the PCR amplification product spanning this sequence by polyacrylamide gel electrophoresis would not be able to readily resolve these two mutations since the products would have the same size. However, these mutations would be distinguished by the HLM. In addition, exon 11 of the CFTR gene contains the sites of many CF mutations including the G542X mutation at base 1756 (see SEQ ID NO 11) and G551D mutation at base 1784 (see SEQ ID NO 16). After a single PCR amplification of the sequence spanning the sites of these mutations, the presence or absence of both mutations can readily be determined by HLM.

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It will be noticed that, although there are some similarities between the HLM technique and ligase chain reaction (LCR), the techniques are, in actuality, very different. LCR itself is an amplification technique that has been known for some time. See, for example, Wu and Wallace, 4 Genomics 560, 1989; also Barany, 88 Proc. Natl. Acad. Sci. USA 189, 1991. In LCR, two portions of oligonucleotides that are each complementary to each chain of a piece of the target gene being amplified (with the two together corresponding to the entire target gene

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portion) are added to the gene sample to be amplified along with a ligase. If the added oligonucleotides complement the target sequence, the ligase will join the two oligonucleotides. LCR is an amplification technique wherein, knowing the sequence to be amplified, it is possible to add to the reaction mixture fragments that are complementary to the target so that, when ligase is added, ligation occurs and the target is amplified. LCR is intended to be repeated for several cycles so that large quantities of the desired sequence can be produced.

HLM, on the other hand, is an analytical technique, wherein probes to one or more suspected sequences which are likely to be found in the target are added to the target. The probes are joined either to a material which can aid separation from the reaction mixture or to a label. Furthermore, the reaction is intended to be run for only one cycle.

There are many variations possible in the HLM procedure. For example, one may combine the detection procedure with column chromatography. One of the oligonucleotide probes in this procedure contains a substituent that will cause it to adhere to a column chromatograph. For example, one of the probes is biotinylated, and the ligated products are separated on avidin-sepharose. The other oligonucleotide might contain a fluorescent marker. Thus, when the sample is passed through a column chromatograph, those oligonucleotides that have been ligated will adhere to the chromatograph and be fluorescent. Thus fluorescence within the column is an indicator that the ligation has occurred. The use of the fluorescence donor/acceptor pair discussed above can also be used in the column chromatography technique.

Another variation deals with the moieties to which the probes are connected. Although in most cases one probe will be connected to a moiety that permits separation of the ligated probes and the second probe will be attached to a label moiety, it is possible that the two probes can be attached to other moieties, such as other sequences. For example, the probes can be connected to the two components of the midivariant sequence, if the QB replicase system is being used. In this case, one probe is connected to one portion of the midivariant sequence (e.g., midivariant A) and the second is connected to the second portion of midivariant sequence (i.e., midivariant B). If replication is observed in the reaction with QB replicase after the ligation step, it is an indication that the two probes were ligated. The fact that the probes were ligated is an indication that the probes have the same sequence as the target.

A further variation of the technique involves the timing for the addition of the flash reagent when using certain luminescent labels, such as acridinium esters. After denaturation and separation of the ligated probes, it is possible to add DNAase before the addition of the flash reagent. This will allow for a more sensitive test, since it has been found that the presence of the insoluble probe interferes at times with the amount of light given off when the flash reagents are added. The separation of the insoluble probe before the addition of the flash reagent thus allows a higher specific signal to be generated.

Another variation concerns the number of probes which are utilized. Although the preferred technique involves the use of two probes, one which aids in separation and

one which aids in detection, it should be noted that more than two probes can be used. In this case, one of the probes contains the separating moiety and one the detection moiety. If the separating moiety is on one terminal probe and the label moiety on the other terminal probe, the probes between them would not need to contain a label. In this experiment, if, after ligation, the label probe is attached to the separating probe, it can be concluded that the intermediate probes also hybridized to the target, for, if this were not the case, the label probe would not be ligated to the probe moiety which includes the separating probe. Another variation of the multiple probe experiment is the one where one probe contained the separating moiety and all the other probes are labeled. In this case, after ligation, the amount of label attached to the separated moiety is an indication of whether all the probes were ligated. The use of more than 2 probes, when combined with the variations discussed above (e.g., analysis with and without ligation, analysis before denaturation, etc.) leads to a number of analytical variations which will be apparent to those with ordinary skill in this area.

Furthermore, the location of the separating or detecting moiety can be varied. Although it is preferred that they be at the terminal end of a probe, it is possible that they be connected to the probe in any location so long as they do not interfere with hybridization and ligation.

A further advantage of the novel technique is that it is now possible to discriminate between two mutations that are closely related to each other (e.g., mutations that occur at adjacent nucleotides).

A further advantage of the instant invention is that different markers can be used on different probes in the same experiment. Thus, for example, the presence of one of two possible mutations can be determined in one test, with each probe using a different marker. Both markers could be separated from the sample and the two could be distinguished by their differing absorption spectra, for example. Furthermore, by using different insoluble particles, the 2 probes can be separated from each other before analysis. For example, if one probe relies on an insoluble non-magnetic particle and the other uses an insoluble magnetic particle, the magnetic field could be applied first in order to remove the magnetic particles and the markers attached thereto, and the remaining solution can be centrifuged to remove the non-magnetic insoluble particles with its attached marker. Other variations of these separation techniques will be apparent to those with skill in the art. Thus, these two markers could still be distinguished from each other even though they both have the same label. By combining in one experiment variations of techniques utilizable for both separation and detection, it is possible to determine the presence of one of several mutations or other genetic variations in the one experiment. For example, magnetic (M) particles could be used on some probes, non-magnetic (NM) on others; some of the second oligonucleotides in the probe could use chemiluminescent material A, while others could use chemiluminescent material B. Thus, by varying just these 2 parameters, 4 mutations could be detected in one experiment. (I.e., where the adducts to the probes are M-A, NM-A, M-B, NM-B) By utilizing particles that can be separated from each other with different markers (i.e., those with different spectral or other characteristics), it would be possible to detect many genetic variations in

one experiment.

A further advantage of the novel procedure is that the technique is more sensitive than previous techniques. This increased sensitivity is due to several factors. For example, the analytical techniques for determining the presence of target material are more sensitive; the solution technique of separating the insoluble particle and analyzing the marker attached thereto is much more sensitive than the process of using electrophoresis to separate the components and to rely on staining to qualitatively and quantitatively determine the presence of the target.

Further variations of the above procedure are possible. For example, after the insoluble particles are separated from the reaction mixture, the quantity of target present can be determined by measuring the amount of marker while the marker is still attached to the insoluble material when the insoluble particles are precipitated (as, for example, using a classical quantitative analysis on insoluble material). Alternatively, after the insoluble particles are separated from the reaction mixture, the particles can be resuspended and the marker determined while on the resuspended particle. Furthermore, the marker can be separated from the insoluble particle and be measured when both the marker and insoluble particle are in solution or suspended. Also, after separation of the marker from the insoluble particle, the insoluble particle can be separated from the solution and the marker can be measured in the absence of the insoluble particle.

In addition to detecting specific DNA sequences, HLM

may also be used to detect specific RNA sequences provided that the ligase used can ligate probes which are hybridized to an RNA target. The technique can also be used to analyze viral materials and other sequences of polynucleic acids. In addition, since the examples illustrate the ability of HLM to distinguish sequences which differ at positions at sites other than the ligation junction of the probes, this method can be readily adapted to scan large segments of sequence even up to whole genes for alterations distinct from a normal sequence.

Further variations of the invention will be apparent to those with ordinary skill in the art. The following examples illustrate various aspects of the invention but are not intended to limit its usefulness.

Example 1: Simultaneous Detection of Normal and Delta F-508 Alleles Using Chemiluminescent Hybridization Ligation Assay with DMAE and LEAE Labeled Probes

A chemiluminescent hybridization-ligation method was tested for its ability to detect the delta F-508 (SEQ ID NO 1) mutation in cystic fibrosis in samples of human DNA amplified by PCR. In addition, the presence of both the normal (SEQ ID NO 2) and delta F-508 alleles was simultaneously determined for each sample by using probes specific for each allele but labeled with two different acridinium ester derivatives (DMAE and LEAE). The DMAE derivative (dimethyl acridinium ester) chemiluminesces at the shorter wavelength range (400-500 nm) and LEAE (longer wavelength emitting acridinium ester) chemiluminesces at the longer wavelength range (500-600 nm).

Nine samples of human DNA (250 ng) obtained from an outside laboratory were amplified by the polymerase chain reaction (see Saiki et al, 239 Science 487, 1988). The primers used were obtained from Genset (Paris, France) and had the following sequences:

C16B (Sequence ID NO 3): 5' GTT TTC CTG GAT TAT GCC TGG CAC 3'

C16D (Sequence ID NO 4): 5' GTT GGC ATG CTT TGA TGA CGC TTC 3'

The target sequence amplified in the PCR reaction with these primers consisted of 97 bp (94 bp for delta F-508 allele) spanning bases 1611-1708 of the CFTR gene (3).

The PCR reactions (75 ul) contained 30 pmol each primer, 1.9 mM MgCl₂, 200 uM each ATP, TTP, GTP, and CTP; and 2.5 U Taq DNA polymerase. After denaturation at 95°C for 5 min, the samples were amplified by 30 cycles of PCR consisting of annealing at 60°C for 45 sec, extension at 72°C for 1 min and denaturation at 95°C for 45 sec. After the last cycle the samples were incubated at 72°C for 5 min.

An aliquot of the PCR reaction solution was denatured and then added to the chemiluminescent detection reaction:

100 ul TE, 4X SSC, 0.1% BSA, 0.02% Tween-20, 5% dextran sulfate containing 10 ug paramagnetic particles (PMP) with the immobilized probe (PMP) and 100 fmol of each acridinium ester (508.CF-DMAE and 508.NOR-LEAE) labeled probe. The sequences of the detection probes were:

PMP.508 (SEQ ID NO 5): 5' CCT AGT CCA AGT ACG GCG CCG AAG
AGG CCC TAT ATT CAT CAT AGG AAA CAC CA 3'

5 508.CF (SEQ ID NO 6): 5' ATG ATA TTT TCT TTA ATG GTG CCA
3'

508.NOR (SEQ ID NO 7): 5' AAG ATG ATA TTT TCT TTA ATG GTG
CCA 3'

10 The possible assay formats are summarized as shown in
Figure 1.

15 The probes were hybridized to the PCR product at 45°C
for 15 minutes. Unhybridized AE probes were removed by
magnetic separation of the particles and decanting the
supernatant. The particles were washed with 2X SSC/0.1%
Tween-20.

20 Hybridized probes were ligated by resuspending
particles in 100 ul 50 mM Tris, pH 7.6, 10 mM MgCl₂, 1 mM
ATP, 1 mM DTT, 5% polyethelyene glycol 8000 and 200 mM
NaCl containing 2 U T4 DNA ligase. Reactions were
25 incubated at 37°C for 15 minutes. After separating and
washing the particles as described, the hybridized but
non-ligated AE probes were dissociated by resuspending the
particles in 150 ul H₂O and incubating at 65°C for 10
minutes. The particles were separated and the supernatant
containing dissociated AE probe removed. The particles
30 were washed once as described and then resuspended in 100
ul 10 mM Tris, pH 7.5, 10 mM MgCl₂, 1 mM EDTA, and 0.05
ug/ul DNase I (BRL). The particle suspension was flashed
using standard flash reagents (see, for example, Law et

al, U.S. Patent No. 5,241,070) and the chemiluminescence detected in a dual wavelength luminometer so that the chemiluminescent signals from the two different labels were measured simultaneously.

5

The results of the chemiluminescent hybridization-ligation assay for PCR amplified product are shown in Figure 2. The chemiluminescent signal obtained clearly identified the presence of the delta F-508 and normal alleles in the PCR amplified products. As expected, the allele specific probes hybridized to each sample irrespective of genotype (data not shown), the subsequent ligation step discriminated between the sequences of the two different alleles since efficient ligation was observed only at the junction of the hybridized probes which were perfectly complementary with the target sequence (see Figure 1).

The chemiluminescent detection of these alleles allowed the diagnosis of these samples which was in complete accord with the analysis of the same samples by an independent laboratory with one exception (See Example 2).

Each sample was analyzed with both delta F-508 and normal allele specific AE probes in order to unambiguously assign the genotype of the samples. The magnitude of the chemiluminescence (Figure 2) also indicated the genotype of the sample in that an intermediate level of chemiluminescence was observed for heterozygous individuals (for example the normal allele in samples 1, 7, and 9) relative to the chemiluminescence of the homozygous cases. Sample 7 was an exceptional sample in that the magnitude of the chemiluminescence from the 508.NOR-LEAE

probe indicated that the sample was heterozygote for this allele, but the signal from the 508.CF-DMAE probe indicated that this sample was negative for the delta F-508 mutation. Taken together these results suggested that the second allele in this sample contained neither the delta F-508 nor the normal sequence but a second cystic fibrosis mutation instead. These PCR products were also analyzed by electrophoresis on an 8M urea/10% polyacrylamide gel and the resolved bands visualized by ethidium bromide staining (data not shown). The products from the normal (97 bp) and CF (94 bp) alleles were clearly resolved on the gel, enabling a diagnosis. In addition, heterozygote samples also contained bands resulting from heteroduplex formation which migrated as apparently larger products. In this regard, on the basis of the electrophoretic analysis, samples 1, 7, and 9 appeared identical and would be assigned as delta F-508/normal heterozygotes. As noted above, on the basis of the chemiluminescent data, although sample 7 was heterozygous for the normal allele, it did not appear to contain the delta F-508 allele. This discrepancy was resolved in Example 2 and illustrated the ability of HLM to provide a more accurate diagnosis compared with standard analytical procedures such as electrophoresis.

Example 2: Discrimination of Delta F-508 and Delta I-507 Mutations by Chemiluminescent Hybridization Ligation

The delta F-508 mutation is the most common mutation in cystic fibrosis (CF), occurring in approximately 68% of the cystic fibrosis chromosomes (3). This mutation is a

deletion of three base pairs in exon 10 of the CFTR gene (3). The sequence of exon 10 surrounding this mutation site is shown in Figure 3. The delta I-507 (SEQ ID NO 8) mutation is a much rarer CF mutation which is also a three
5 base pair deletion that partially overlaps the delta F-508 mutation (Figure 3). The sequences of these two CF alleles differ by a single base.

The ability of the chemiluminescent hybridization
10 ligation assay to distinguish the delta F-508 and normal alleles was demonstrated in Example 1. The ability of the assay to distinguish the delta F-508 and delta I-507 alleles was shown in this example. This application requires that the ligation step distinguish sequences
15 which differ at a single position. In addition, the site of the mismatch in these hybrids occurs at one base removed from the ligation junction. The hybrids formed between the delta F-508 and delta I-507 probe sets and the different target sequences are shown in Figure 4.

20

The same nine clinical samples of human DNA which had been amplified by the polymerase chain reaction (example 1) were analyzed for the presence of the delta I-507 allele. In this assay, a similar format was followed as
25 in example 1 with the exception that the solid phase and acridinium ester (DMAE) labeled probes were as follows:

PMP.507 (SEQ ID NO 9) : 5' CCT AGT CCA AGT ACG GCG CCG
AAG AGG CCC TAT ATT CAT CAT AGG AAA CAC CAA AG 3'
30 507.CF (SEQ ID NO 10): 5' ATA TTT TCT TTA ATG GTG CCA GGC
3'

An additional three samples (30, 31, and 32) of PCR

amplified human DNA were assayed for the delta F-508, delta I-507, and normal alleles using the same protocol as well as a protocol utilizing Taq DNA ligase (Epicentre Technologies). The thermostablility of Taq DNA ligase permitted the ligation reaction to be carried out at higher temperature as well as allowing the hybridization and ligation steps to be carried out simultaneously. The buffer for the simultaneous hybridization-ligation consisted of 20 mM Tris, pH 8.3, 200 mM KCl, 10 uM tRNA, 10 mM MgCl₂, 0.5 mM NAD, and 0,01% Triton X-100. The reactions contained 100 units Taq DNA ligase. The simultaneous hybridization-ligations were carried out at 60°C for 30 minutes. The remainder of the protocol was identical to that for the assays employing T4 DNA ligase.

The results of the chemiluminescent hybridization ligation assay of the nine samples are shown in Figure 5. Only sample 7 was positive for the delta I-507 allele. These results taken together with those of Example 1 permit the following assignments for these samples:

Table I
GENOTYPE OF CLINICAL SAMPLES

1	2	3	4	5	6	7	8	9
508/N	508/508	N/N	N/N	N/N	508/508	507/N	508/508	508/N

The results of the three additional samples assayed for the delta F-508, delta I-507, and normal alleles are shown in Figures 6 and 7. The assignments made for these samples were confirmed by sequencing.

The results shown in examples 1 and 2 illustrate the

ability of chemiluminescent hybridization-ligation assay to discriminate sequences differing by a single base even when the site of this difference occurs one base removed from the site of the ligation junction. In this particular application, the cystic fibrosis mutations delta F-508 and delta I-507 were distinguished. This enabled samples that had been previously characterized as delta F-508/N to be correctly assigned as delta I-507/N. The assignment of samples 7 and 30 as delta F-508/N heterozygotes had been made on the basis of the electrophoretic mobility of the PCR products of these samples. But analysis of these PCR products by polyacrylamide gel electrophoresis fails to readily distinguish the delta F-508 and delta I-507 mutations, since both mutations consist of three base pair deletions, the PCR products from these alleles are the same size.

Example 3: Assay for the G542X Cystic Fibrosis Mutation

The previous examples have demonstrated the ability of the chemiluminescent hybridization-ligation assay to distinguish the normal and cystic fibrosis alleles at the sites of the delta F-508 and delta I-507 mutations. A third cystic fibrosis mutation, G542X (SEQ ID NO 11), is a point mutation occurring in exon 11 of the CFTR gene. The current method for detecting this mutation requires sequencing, which is a lengthy and laborious procedure. The chemiluminescent hybridization-ligation assay to detect this mutation must be able to distinguish the single base substitution which differs between the normal (SEQ ID NO 12) and cystic fibrosis alleles.

The same simultaneous hybridization-ligation assay using Taq DNA ligase as described in Example 2 was used in

the G542X assay. The sequences of the G542X probes were:
PMP.G542X (SEQ ID NO 13): 5' CCT AGT CCA AGT ACG GCG CCG
AAG AGG CCA CTC AGT GTG ATT CCA CCT TCT C 3'

5 G542X.CF (SEQ ID NO 14): 5' AAA GAA CTA TAT TGT CTT TCT
CTG CAA 3'

G542X.NOR (SEQ ID NO 15): 5'CAA GAA CTA TAT TGT CTT TCT
CTG CAA 3'

10

The results of the assay are shown in Figure 8 and indicate
that the only G542X.CF probe was ligated with the G542X
sequence and only the G542X.NOR probe was ligated with the
normal sequence. In this example, T-C and G-A mismatches
15 at the ligation junction were not efficiently ligated.

Example 4: Simultaneous Assay for Delta F-508 and G542X

20 The chemiluminescent hybridization-ligation assay can
be used for the simultaneous detection of multiple
sequences. One illustration of this capability was the
simultaneous detection of the delta F-508 and normal
alleles in a single assay (Example 1). Another application
is the detection of two or more mutations which underly an
25 inherited disease or cancer. For example, more than 200
mutations have been described which underly cystic
fibrosis. The delta F-508 mutation is the most common one,
occurring in approximately 68% of the cystic fibrosis
chromosomes. The second most frequent cystic fibrosis
30 mutation is the G542X mutation, a point mutation occurring
in exon 11 of the CFTR gene. Instead of using the two
different acridinium ester labels to simultaneously detect
the normal and CF alleles at a single locus, the delta F-

508 and G542X mutations may be detected in a single assay. In principle, as many mutations as there are acridinium ester derivatives with distinct chemiluminescent properties, may be detected in a single assay. Alternative labels, such as fluorophores, may permit a still greater number of loci to be simultaneously detected.

A model assay to test feasibility was performed with synthetic target sequences. The assay protocol was the same as that described in Example 2 using Taq DNA ligase and simultaneous hybridization-ligation. Two solid phases and two acridinium ester labeled probes, one set each for the delta F-508 and G542X mutations were used. The probes for the the delta F-508 mutation were the same as those described in Example 1 with the exception that the acridinium ester label was LEAE. The sequences of the G542X probes were the same as those used in example 3. The results of the assay are summarized in Figure 9 and show that the delta F-508 and G542X sequences were detected with whatever combinations of targets were employed. This demonstrates the feasibility of detecting multiple genetic mutations in the same assay.

Example 5: Discrimination of G551D, G551S, and Q552X Cystic Fibrosis Mutations.

Exon 11 of the CFTR gene contains the sites for many other cystic fibrosis mutations in addition to the G542X mutation described in the examples above. The presence of multiple mutation sites in the relatively short span of sequence of exon 11 has heretofore resulted in the necessity of sequencing the PCR product from this exon in order to detect and discriminate these possible mutations.

The ability of HLM to simplify analysis of exon 11 cystic fibrosis mutations required that the specificity of this method enable discrimination of closely clustered mutation sites. The G551D mutation (SEQ ID NO 16) is one of the more common cystic fibrosis mutations, accounting for approximately 0.5% of the observed frequency, this is a point mutation in which G1784 in the normal gene (SEQ ID NO 17) is changed into an A. Close to the site of the G551D are the G551S (SEQ ID NO 18) at base 1783, and Q552X (SEQ ID NO 19) at base 178. In addition the nature of the G551D mutation requires the discrimination of a G-T mismatch by HLM, one of the most difficult mismatches to discriminate (see below). The ability of HLM to detect the G551D mutation and discriminate between it and the other mutation sites near to it was demonstrated in this example.

The sequences of the probes used in the G551D assay were as follows:

PMP.G551D (SEQ ID NO 20): CCT AGT CCA AGT ACG GCG CCG AAG
AGG CCC TAA AGA AAT TCT TGC TCG TTG A

G551D.CF (SEQ ID NO 21): TC TCC ACT CAG TGT GAT TCC AC

G551D.NOR (SEQ ID NO 22): CC TCC ACT CAG TGT GAT TCC AC

In these assays, G551D.CF and G551D.NOR were labeled with ³²P at their 5' termini. Detection of ligation product was accomplished by liquid scintillation counting. Assays were performed using either the standard T4 DNA ligase and Taq DNA ligase protocols described above as well as modifications to these protocols by altering the salt

5 conditions in order to improve specificity of HLM. For T4 DNA ligase, this involved increasing the NaCl concentration from 200 to 600 mM. The Taq DNA ligase protocol was altered by substituting NaCl for KCl. The exact ligation conditions are indicated with the figures below.

10 The results of HLM analysis using the T4 DNA ligase protocol with 200 mM NaCl are shown in Figure 10. Although HLM discriminated between the G551D sequence and the sequences for the other mutations, there was essentially no discrimination between the G551D sequence and the normal sequence with the G551D.CF probe. The specificity for HLM using T4 DNA ligase was improved by increasing the NaCl concentration to 600 mM (Figure 11). Even better discrimination was obtained by employing HLM with Taq DNA ligase (Figure 12), under these conditions the different sequences were readily discriminated with either G551D.CF or G551D.NOR probes. Using the the Taq DNA ligase protocol with 200 mM KCl and G551D.CF, discrimination was readily apparent between the G551D and normal sequences, but some ligation above background was observed with the normal target. This signal was able to be depressed further by substitution of NaCl into the Taq DNA ligase protocol (Figure 13).

25

Taken together these results indicate the ability to HLM to discriminate sequences which differ by only a single base, even when the site of the single base change occurs at positions removed from the ligation junction.

30

Example 6: Systematic Evaluation of the Specificity of Hybridization-Ligation Utilizing a p53 Model.

The preceding examples have establish the ability of

HLM to discriminate sequences with subtle differences including deletions, insertions, and point mutations. In each of the cases examined conditions were established which permitted the specificity of ligation to discriminate these sequence differences. In this example, the specificity of HLM is systematically evaluated by testing its ability to discriminate sequences which differ by all of the possible combinations of mismatches.

The p53 gene codes for a protein which functions as a tumor suppressor. Mutations in this gene are observed in a wide variety of tumors, the most frequent positions for these mutations cluster about codons 175, 245, and 248. A portion of the p53 gene surrounding codon 175 (SEQ ID NO 23) was chosen as the model for the systematic evaluation of the specificity of ligation. The target and probe sequences for this model are shown in Figure 14. Hybridization and ligations were carried out in solution. For assays utilizing T4 DNA ligase, probes (³²P-SEQ ID NO 24 and 25) were mixed with targets in ligation buffer containing the indicated NaCl concentration. Reactions were incubated at 37°C for 15 min, T4 DNA ligase (1 U) added and reactions incubated at 37°C for a further 15 min. An aliquot of the reactions was analyzed by denaturing (8 M urea) polyacrylamide (15%) gel electrophoresis. Bands corresponding to the ligation product and the unligated oligomer were excised and counted by liquid scintillation counting, and the per cent of the total oligomer ligated calculated. The assays utilizing Taq DNA ligase were carried out similarly except that the ligase was added at the start of the reactions.

The ability of T4 DNA ligase to discriminate mismatches occurring at the 5' phosphate or 3' OH

nucleotide at the ligation junction in 200 mM NaCl is summarized in Table II. Mismatches at the 3' OH nucleotide were much more easily discriminated than mismatches at the 5' P nucleotide. Increasing the NaCl concentration to 600 mM improved the specificity of T4 DNA ligase over that observed at 200 mM. At 600 mM NaCl, all possible mismatches at either the 5' P or 3' OH positions were discriminated, even the G-T and C-A mismatches (Table III).

10 The specificity of Taq DNA ligase at 45°C in 200 mM KCl was better than that of T4 DNA ligase (Table IV). Inclusion of NaCl in the Taq ligase buffer also improved the specificity of Taq DNA ligase somewhat (Table V).

TABLE II
SPECIFICITY OF T4 DNA LIGASE IN 200 mM NaCl^a

	TARGET BASE	3' OH PROBE BASE				5' P PROBE BASE			
		C	T	A	G	C	T	A	G
5									
10	G	80	51	1.9	9.8	52	41	15	31
	A	20	72	1.4	3.6	58	65	45	15
15	T	2.2	4.2	77	8.7	56	54	62	64
	C	1.4	11	53	69	10	21	29	40
20									

^aThe results summarized in the table represent the percentages of the total amount of the limiting probe which was ligated as determined by PAGE analysis. The table is arranged so that the complementary pairs of nucleotides fall on the diagonal, the off-diagonal entries are the possible combinations of mismatches. Refer to Figure 14 for the sequence of the hybrids formed between probes and targets.

TABLE III
SPECIFICITY OF T4 DNA LIGASE IN 600 mM NaCl^a

	TARGET BASE	3' OH PROBE BASE				5' P PROBE BASE			
		C	T	A	G	C	T	A	G
5									
10	G	54	1.1	0.7	1.5	36	17	1.4	0.7
	A	2.2	64	1.0	3.6	3.0	31	1.3	1.0
15	T	1.1	1.3	55	2.0	1.6	14	43	37
	C	1.1	1.1	1.1	44	0.6	2.0	2.5	25
20									

^aThe results summarized in the table represent the percentages of the total amount of the limiting probe which was ligated as determined by PAGE analysis. The table is arranged so that the complementary pairs of nucleotides fall on the diagonal, the off-diagonal entries are the possible combinations of mismatches. Refer to Figure 14 for the sequence of the hybridis formed between probes and targets.

TABLE IV

SPECIFICITY OF Taq DNA LIGASE IN 200 mM KCl^a

	TARGET BASE	3' OH PROBE BASE				5' P PROBE BASE			
		C	T	A	G	C	T	A	G
	G	60	8.2	0.6	1.8	34	14	0.6	1.9
15	A	0.8	64	1.1	2.6	7.0	43	5.0	3.0
	T	1.5	2.7	71	3.3	5.0	16	43	11
20	C	2.7	2.0	1.6	61	2.0	4.0	11	34

25 ^aThe results summarized in the table represent the percentages of the total amount of the limiting probe which was ligated as determined by PAGE analysis. The table is arranged so that the complementary pairs of nucleotides fall on the diagonal, the off-diagonal entries are the possible combinations of mismatches. Refer to Figure 14

30 for the sequence of the hybridis formed between probes and targets.

TABLE V

SPECIFICITY OF Taq DNA LIGASE IN 25 mM KCl/75 mM NaCl^a

5	TARGET BASE	3' OH PROBE BASE			
10		C	T	A	G
	G	31	0.6	0.7	0.7
15	A	1.5	46	0.7	2.5
	T	0.8	2.5	51	1.7
20	C	0.8	0.8	0.8	39

25 ^aThe results summarized in the table represent the
 percentages of the total amount of the limiting probe which
 was ligated as determined by PAGE analysis. The table is
 arranged so that the complementary pairs of nucleotides
 fall on the diagonal, the off-diagonal entries are the
 possible combinations of mismatches. Refer to Figure 14
 30 for the sequence of the hybrids formed between probes and
 targets.

Example 7: Assays for the delta F-508 and Normal Alleles
in PCR Amplified Human DNA Using Percent Ligation as
Diagnostic Criterion

5 Samples of PCR amplified human DNA were received from
an independent laboratory. HLM analysis of the delta F-508
(SEQ ID NO 1) and normal (SEQ ID NO 2) alleles was
performed as described in the previous examples. At the
denaturation step to remove the hybridized but unligated
10 probe, the supernatant containing this released probe was
reserved and flashed separately, in addition to flashing
the PMP which contained the hybridized and ligated labeled
probe. The sum of the chemiluminescent signals from the
supernatant and PMP provides a measure of the total amount
15 of labeled probe which hybridized to the sample. This in
turn provides a measure of the total amount of sample DNA
in the reaction. In addition, the chemiluminescent signal
from the PMP divided by the sum of the chemiluminescent
signals from the supernatant and PMP provides a measure of
20 the fraction of the hybridized probe which was ligated.
Since the amount of PCR product obtained from each sample
of human DNA may vary from sample to sample, the fraction
of labeled probe ligated provides a clearer distinction
between samples which are homozygous for an allele, samples
25 which are heterozygous for an allele, and those which do
not contain the allele. In addition, since the probes may
be expected to hybridize to delta F-508, normal, and delta
I-507 alleles, determining the chemiluminescence of the
labeled probe released at the denaturation step, provides
30 a means to determine that the assay components are
functioning properly for those samples which are ligation
negative.

The HLM results for the delta F-508 assay calculated

as per cent ligation are summarized in Figure 15. A clear discrimination between homozygous, heterozygous, and negative samples could be made based upon the calculated percentage of the labeled probe ligated. This calculated parameter turns out to be a more reliable diagnostic index than the raw chemiluminescent data because the samples were found to vary more than 5-fold in the total amount of DNA present (data not shown). In Figure 16, similar HLM results for the assay for the normal allele are compared with those of the delta F-508 assay.

Example 8: Effect of NaCl Concentration on the Discrimination of Delta F-508 and Delta I-507 Alleles

The sequences for these two alleles differ at a single position. (See Figure 4). Using HLM with T4 DNA ligase in 200 mM NaCl, it was possible to discriminate between these two cystic fibrosis mutations (See Examples 1 and 2). In light of the results in Example 6 showing the effect of NaCl concentration on the specificity of T4 DNA ligase with the p53 sequences, the effect of NaCl concentration on the ability of the T4 DNA ligase to discriminate between the delta F-508 and delta I-507 sequences using the delta F-508 probes was examined. HLM was performed on delta F-508 and delta I-507 synthetic target sequences as described except that ligation buffers were made up with either 200, 400, 600, 800, or 1000 mM NaCl. At the denaturation step, the supernatant was reserved and flashed separately in order to determine the amount of hybridized but unligated probe. The percentage of the 508.CF-DMAE probe which had been ligated out of the total amount hybridized was calculated as described in Example 7.

The results are summarized in Figure 17. There was sufficient discrimination by T4 DNA ligase in 200 mM NaCl to distinguish between the delta F-508 and delta I-507 sequences. Increasing the salt concentration up to 600 mM improved the discrimination between these sequences by suppressing the amount of ligation observed with the delta I-507 sequence while maintaining the level of ligation with the delta F-508 sequence. At NaCl concentrations above 600 mM, the ligation with the delta F-508 target begins to decline.

Example 9. Hybridization-Ligation Assay for Δ F-508 with Biotinylated Probe and Avidin-PMP.

Hybridization and ligation reactions were carried out in solution with a biotinylated probe (Biotin-CF1) and an acridinium ester labeled probe (508.CF-AE). Hybridization-ligation reactions were carried out in 100 μ l buffer (20 mM Tris, pH 8.3, 100 mM NaCl, 100 mM KCl, 10 mM MgCl₂, 10 μ M t RNA, 0.5 mM NAD and 0.01% Triton X-100) containing 1 pmol Biotin-CF1, 100 fmol 508.CF-AE, and 100 units Taq DNA ligase at 50°C. Reactions were initiated by the addition of either 1) 1 fmol normal target, 2) 1 fmol Δ I-507 target, 3) 1 fmol Δ F-508 target or 4) No target. Reactions were incubated at 50°C for 1 hour. To each reaction was added 10 μ g avidin-PMP (Promega) and incubated an additional 10 minutes. The particles were separated magnetically, the supernatant aspirated, and the particles washed with 0.2X SSC/0.1% Tween-20. The washes were repeated twice. The avidin-PMP were resuspended in 150 μ l wash buffer and incubated at 55°C for 10 minutes to remove hybridized but unligated probe. The avidin-PMP were separated magnetically, the supernatant removed and

flushed. The avidin-PMP were washed once, resuspended in 100 μ l 10 mM MgCl₂, 50 mM Tris, pH 7.5 containing DNase I (BRL) and flashed. The results of the assay are summarized in the table below.

5

	<u>Target</u>	<u>Percent Ligation</u>
	Normal	0%
10	Δ I-507	24%
	Δ F-508	60%

15

This assay format shows the feasibility of the use of a universal solid phase reagent (avidin-PMP) with a biotinylated probe. This format may be especially useful in multiplexing schemes where it is expected that assays for a number of genetic loci may be performed in the same tube. In addition, solution phase hybridizations and ligations may proceed more rapidly than those involving one probe immobilized on PMP.

20

Example 10: Ligation Specificity and G551D Model Results

25

These results extend those given in Example 5 in the instant application. Hybridization-ligation assays were carried out with T4 DNA and Taq DNA ligases as described previously. The results of these assays using the AE probes specific for the G551D sequence and the normal sequence are summarized in the table. Inclusion of the target sequence designated R553X provides additional insight into ligation specificity. (See

30

Table VI.)

In the assay with the G551D.NOR probe, the G551S and Q552X sequences were discriminated. This illustrates the ability of T4 and Taq DNA ligases to discriminate mismatches at positions other than at the ligation junction. Moreover, in the assay with G551D.NOR, T4 DNA ligase appears to be able to discriminate the R553X sequence. This requires the enzyme to discriminate a mismatch 5 bases from the 3' hydroxyl side of the ligation junction.

The results of this assay illustrate the improvement in discrimination one can obtain when two mismatches are present as opposed to a single mismatch. For example, in the assay with G551D.NOR, the R553X sequence is not discriminated by Taq ligase. Viewed in a different way, the 3'(5) T-G mismatch does not interfere with the ligation by Taq DNA ligase. In the assay with G551D.CF, the level of misligation for the Normal target sequence was 5% while that for R553X was 2%. This illustrates the utility of introducing a mismatch at a site several bases away from the ligation junction in improving discrimination. Ultimately such a strategy may permit discrimination of mismatches not currently discriminated. This aspect may be illustrated by considering the assay with G551D.CF and T4 DNA ligase. This enzyme did not discriminate the G-T mismatch at 5'(1). But the combination of mismatches that occurs in R553X was discriminated.

30

TABLE VI
LIGATION SPECIFICITY IN THE G551D ASSAY

	Probes	Target	Mismatch	Per Cent Ligation	
				T4 ¹	Taq ²
5					
	G551D.CF	NOR	5' (1) G-T	27%	5%
		G551D		30%	45%
10		G551S	5' (1) G-T 5' (2) A-C	1.0%	0%
		Q552X	5' (1) G-T 3' (2) T-G	1.5%	0.3%
15		R553X	5' (1) G-T 3' (5) T-G	4%	2%
	G551D.NOR	NOR		43%	53%
20		G551D	5' (1) A-C	5.2%	8%
		G551S	5' (2) A-C	5.4%	4%
		Q552X	3' (2) T-G	1.9%	2%
		R553X	3' (5) T-G	10%	51%
25	¹ 600 mM NaCl				
	² 100 mM KCl/100 mM NaCl, 55°C				

Example 11: Discrimination of Mismatches at Positions
Away from the Ligation Junction

A systematic evaluation of the abilities of T4 and
5 Taq DNA ligases to discriminate mismatches at positions
one base removed from the ligation junction was
undertaken with the p53 model described in the patent
application. These positions are designated as 5'(2)
and 3'(2) depending on the side of the ligation
10 junction. The results are summarized in the enclosed
figure. All mismatches were discriminated by both
enzymes at the 3'(2) position. Not all 5'(2) mismatches
were discriminated by either enzyme. (See Figure 18.)

15 Example 12: Effect of Enzyme Concentration on Ligation
Specificity

Two different T4 DNA ligase concentrations were
compared: 1 nM vs 240 nM. The ability of T4 DNA ligase
20 to discriminate 5'(1) and 3'(1) mismatches in the p53
model was evaluated. The results are summarized in the
Figure. Clearly, the ligation specificity for 3'(1)
misimatches improved at the lower enzyme concentration
without significant loss of ligation for the
25 complementary matches. There was also improvement for
the 5'(1) mismatches, especially for the purine-purine

mismatches, but not as significant overall as for the 3' (1) mismatches. (See Figures 19 and 20.)

5

SEQUENCE LISTING

(1) GENERAL INFORMATION

10

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(v) COMPUTER READABLE FORM:

35

(A) MEDIUM TYPE: Diskette-3.50 inch, 1.44Mb storage

(B) IBM Compatible

40

(C) OPERATING SYSTEM: IBM DOS 5.0

(D) SOFTWARE: WORD 6.0

(vi) CURRENT APPLICATION DATA:

45

(A) APPLICATION NUMBER: Not available

(B) FILING DATE: Not available

50

(C) CLASSIFICATION: Not available

5

(vii) PRIOR APPLICATION DATA:**(A) APPLICATION NUMBER: US 08/222,613**

10

(B) FILING DATE: 04-04-1994**(C) CLASSIFICATION: Not available**

15

(viii) ATTORNEY/AGENT INFORMATION:**(A) NAME: Morgenstern, A. S.**

20

(B) REGISTRATION NUMBER: 28,244**(C) REFERENCE/DOCKET NUMBER: CCD-113****(ix) TELECOMMUNICATION INFORMATION:**

25

(A) TELEPHONE: 508-359-3836**(B) TELEFAX: 508-359-3885**

30

(2) INFORMATION FOR SEQ ID NO: 1:**(i) SEQUENCE CHARACTERISTICS:**

35

(A) LENGTH: 47 bases**(B) TYPE: Nucleic acid****(C) STRANDEDNESS: Single**

40

(D) TOPOLOGY: Linear**(ii) MOLECULE TYPE: Genomic DNA**

45

(A) DESCRIPTION: Delta F508, a portion of the sequence of exon 10 of the CFTR gene surrounding base number 1652 with base 1653-1655 deleted.**(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 1:**

50

5 GGC ACC ATT AAA GAA AAT ATC ATT GGT GTT TCC TAT GAT GAA TAT AG 47

(3) INFORMATION FOR SEQ ID NO: 2:

(i) SEQUENCE CHARACTERISTICS:

10

(A) LENGTH: 50 bases

(B) TYPE: Nucleic acid

15

(C) STRANDEDNESS: Single

(D) TOPOLOGY: Linear

(ii) MOLECULE TYPE: Genomic DNA

20

(A) DESCRIPTION: Normal, a portion of the sequence of
exon 10 of the CFTR gene surrounding base number 1652

25

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 2:

GGC ACC ATT AAA GAA AAT ATC ATC TTT GGT GTT TCC TAT GAT GAA 45

TAT AG 50

30

(4) INFORMATION FOR SEQ ID NO: 3:

(i) SEQUENCE CHARACTERISTICS

35

(A) LENGTH: 24 bases

(B) TYPE: Nucleic acid

(C) STRANDEDNESS: Single

40

(D) TOPOLOGY: Linear

(ii) MOLECULE TYPE: Genomic DNA

45

(A) DESCRIPTION: C16B, bases 1611 - 1634 of exon 10 of
the CFTR gene

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 3:

50

5 G TTT TCC TGG ATT ATG CCT GGC AC 24

(5) INFORMATION FOR SEQ ID NO: 4:

(i) SEQUENCE CHARACTERISTICS:

10

(A) LENGTH: 24 bases

(B) TYPE: Nucleic acid

15

(C) STRANDEDNESS: Single

(D) TOPOLOGY: Linear

(ii) MOLECULE TYPE: Genomic DNA

20

(A) DESCRIPTION: C16D, bases 1708 - 1684 of exon 10 of
the CFTR gene

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 4:

25

GTT GGC ATG CTT TGA TGA CGC TTC 24

(6) INFORMATION FOR SEQ ID NO: 5:

30

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 53 bases

35

(B) TYPE: Nucleic acid

(C) STRANDEDNESS: Single

(D) TOPOLOGY: Linear

40

(ii) MOLECULE TYPE: Other DNA/Genomic DNA

(A) DESCRIPTION: PMP.508, bases 1 - 29 is a spacer of
synthetic DNA; bases 30 - 53 consists of bases 1656-
1678 of exon 10 of the CFTR gene

45

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 5:

CCTAGTCCAA GTACGGCGCC GAAGAGGCC CT ATA TTC ATC ATA GGA 46

50

5 AAC ACC A 53

(7) INFORMATION FOR SEQ ID NO: 6:

(i) SEQUENCE CHARACTERISTICS:

10

(A) LENGTH: 24 bases

(B) TYPE: Nucleic acid

15

(C) STRANDEDNESS: Single

(D) TOPOLOGY: Linear

(ii) MOLECULE TYPE: Genomic DNA

20

(A) DESCRIPTION: 508.CF, bases 1629-1652 of exon 10 of
the CFTR gene

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 6:

25

AT GAT ATT TTC TTT AAT GGT GCC A 24

(8) INFORMATION FOR SEQ ID NO: 7:

30

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 27 bases

35

(B) TYPE: Nucleic acid

(C) STRANDEDNESS: Single

(D) TOPOLOGY: Linear

40

(ii) MOLECULE TYPE: Genomic DNA

(A) DESCRIPTION: 508.NOR, bases 1629-1655 of exon 10 of
the CFTR gene

45

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 7:

AA GAT GAT ATT TTC TTT AAT GGT GCC A 27

50 (9) INFORMATION FOR SEQ ID NO: 8:

5

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 47 bases

10

(B) TYPE: Nucleic acid

(C) STRANDEDNESS: Single

(D) TOPOLOGY: Linear

15

(ii) MOLECULE TYPE: Genomic DNA

(A) DESCRIPTION: Delta I-507, a portion of the sequence
of exon 10 of the CFTR gene surrounding base number
1652 with base 1652-1654 deleted.

20

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 8:

GGC ACC ATT AAA GAA AAT ATC TTT GGT GTT TCC TAT GAT GAA 42

25

TAT AG 47

(10) INFORMATION FOR SEQ ID NO: 9:

30

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 56 bases

(B) TYPE: Nucleic acid

35

(C) STRANDEDNESS: Single

(D) TOPOLOGY: Linear

40

(ii) MOLECULE TYPE: Genomic DNA / other DNA

(A) DESCRIPTION: PMP.507: bases 1 - 29 is a spacer of
synthetic DNA; bases 30 - 56 consists of bases
1679 - 1653 of exon 10 of the CFTR gene

45

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 9:

CCTAGTCCAA GTACGGCGCC GAAGAGGCC CT ATA TTC ATC ATA GGA 46

50

AAC ACC AAA G 56

5

(11) INFORMATION FOR SEQ ID NO: 10:**(i) SEQUENCE CHARACTERISTICS:**

10

(A) LENGTH: 24 bases**(B) TYPE: Nucleic acid****(C) STRANDEDNESS: Single**

15

(D) TOPOLOGY: Linear**(ii) MOLECULE TYPE: Genomic DNA**

20

(A) DESCRIPTION: 507.CF, consists of bases 1626-1649 of exon 10 of the CFTR gene**(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 10:**

25

AT ATT TTC TTT AAT GGT GCC AGG C 24**(12) INFORMATION FOR SEQ ID NO: 11:****(i) SEQUENCE CHARACTERISTICS:**

30

(A) LENGTH: 54 bases**(B) TYPE: Nucleic acid**

35

(C) STRANDEDNESS: Single**(D) TOPOLOGY: Linear****(ii) MOLECULE TYPE: Genomic DNA**

40

(A) DESCRIPTION: G542X, bases 1731 - 1784 of exon 11 of the CFTR gene with T substituted for G at base 1756**(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 11:**

45

**T GCA GAG AAA GAC AAT ATA GTT CTT TGA GAA GGT GGA ATC ACA CTG 46
AGT GGA GG 54****50 (13) INFORMATION FOR SEQ ID NO: 12:**

5

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 54 bases

10

(B) TYPE: Nucleic acid

(C) STRANDEDNESS: Single

(D) TOPOLOGY: Linear

15

(ii) MOLECULE TYPE: Genomic DNA

(A) DESCRIPTION: NOR1731.54, bases 1731 - 1784 of exon 11
of the CFTR gene

20

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 12:

T GCA GAG AAA GAC AAT ATA GTT CTT GGA GAA GGT GGA ATC ACA CTG 46

25

AGT GGA GG 54

(14) INFORMATION FOR SEQ ID NO: 13:

(i) SEQUENCE CHARACTERISTICS:

30

(A) LENGTH: 52 bases

(B) TYPE: Nucleic acid

35

(C) STRANDEDNESS: Single

(D) TOPOLOGY: Linear

(ii) MOLECULE TYPE: Other DNA/Genomic DNA

40

(A) DESCRIPTION: PMP.G542X, bases 1-27 consists of a
spacer of synthetic DNA, bases 28 - 52 consists of
bases 1781-1757 of exon 11 of the CFTR gene

45

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 13:

CCTAGTCCAA GTACGGCGCC GAAGAGGCC ACT CAG TGT GAT TCC ACC 47

TTC TC 52

50

5 (15) INFORMATION FOR SEQ ID NO: 14:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 27 bases

10

(B) TYPE: Nucleic acid

(C) STRANDEDNESS: Single

15

(D) TOPOLOGY: Linear

(ii) MOLECULE TYPE: Genomic DNA

(A) DESCRIPTION: G542X.CF, bases 1756 - 1730 of exon 11
of the CFTR gene with T substituted for C at base
1756

20

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 14:

25 A AAG AAC TAT ATT GTC TTT CTC TGC AA 27

(16) INFORMATION FOR SEQ ID NO: 15:

(i) SEQUENCE CHARACTERISTICS:

30

(A) LENGTH: 27 bases

(B) TYPE: Nucleic acid

35

(C) STRANDEDNESS: Single

(D) TOPOLOGY: Linear

(ii) MOLECULE TYPE: Genomic DNA

40

(A) DESCRIPTION: G542X.NOR, bases 1756 - 1730 of exon 11
of the CFTR gene

45 (xi) SEQUENCE DESCRIPTION: SEQ ID NO: 15:

CAA GAA CTA TAT TGT CTT TCT CTG CAA 27

(17) INFORMATION FOR SEQ ID NO: 16:

50

5 (i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 45 bases

(B) TYPE: Nucleic acid

10

(C) STRANDEDNESS: Single

(D) TOPOLOGY: Linear

15 (ii) MOLECULE TYPE: Genomic DNA

(A) DESCRIPTION: G551D, bases 1763 - 1807 of exon 11 of
the CFTR gene with A substituted for the G at base
1784

20

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 16:

GT GGA ATC ACA CTG AGT GGA GAT CAA CGA GCA AGA ATT TCT TTA G 45

25 (18) INFORMATION FOR SEQ ID NO: 17:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 45 bases

30

(B) TYPE: Nucleic acid

(C) STRANDEDNESS: Single

35

(D) TOPOLOGY: Linear

(ii) MOLECULE TYPE: Genomic DNA

(A) DESCRIPTION: NOR17632.45, bases 1763 - 1807 of exon
11 of the CFTR gene

40

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 17:

GT GGA ATC ACA CTG AGT GGA GGT CAA CGA GCA AGA ATT TCT TTA G 45

45

(19) INFORMATION FOR SEQ ID NO: 18:

(i) SEQUENCE CHARACTERISTICS:

50

5 (A) LENGTH: 45 bases

(B) TYPE: Nucleic acid

(C) STRANDEDNESS: Single

10

(D) TOPOLOGY: Linear

(ii) MOLECULE TYPE: Genomic DNA

15 (A) DESCRIPTION: G551S, bases 1763 - 1807 of exon 11 of
the CFTR gene with A substituted for G at base 1783

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 18:

20 GT GGA ATC ACA CTG AGT GGA AGT CAA CGA GCA AGA ATT TCT TTA G 45

(20) INFORMATION FOR SEQ ID NO: 19:

25 (i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 45 bases

(B) TYPE: Nucleic Acid

30

(C) STRANDEDNESS: Single

(D) TOPOLOGY: Linear

35 (ii) MOLECULE TYPE: Genomic DNA

(A) DESCRIPTION: Q552X, bases 1763 - 1807 of exon 11 of
the CFTR gene with T substituted for C at base 1786

40 (xi) SEQUENCE DESCRIPTION: SEQ ID NO: 19:

GT GGA ATC ACA CTG AGT GGA GGT TAA CGA GCA AGA ATT TCT TTA G 45

(21) INFORMATION FOR SEQ ID NO: 20:

45

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 52 bases

50

(B) TYPE: Nucleic acid

5

(C) STRANDEDNESS: Single

(D) TOPOLOGY: Linear

10

(ii) MOLECULE TYPE: Other DNA/Genomic DNA

(A) DESCRIPTION: PMP.G551D, bases 1-29 consist of a
spacer of synthetic DNA, bases 30-52 consist of bases
1785 - 1807 of exon 11 of the CFTR gene

15

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 20:

CCTAGTCCAA GTACGGCGCC GAAGAGGCC C TAA AGA AAT TCT TGC 45

20

TCG TTG A 52

(22) INFORMATION FOR SEQ ID NO: 21:

25

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 22 bases

(B) TYPE: Nucleic acid

30

(C) STRANDEDNESS: Single

(D) TOPOLOGY: Linear

35

(ii) MOLECULE TYPE: Genomic DNA

(A) DESCRIPTION: G551D.CF, bases 1784 - 1763 of exon 11
of the CFTR gene with T substituted for C at base
1784

40

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 21:

TC TCC ACT CAG TGT GAT TCC AC 22

45

(23) INFORMATION FOR SEQ ID NO: 22:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 22 bases

50

(B) TYPE: Nucleic acid

5

(C) STRANDEDNESS: Single

(D) TOPOLOGY: Linear

10

(ii) MOLECULE TYPE: Genomic DNA

(A) DESCRIPTION: G551D.NOR, bases 1784-1763 of exon 11 of the CFTR gene

15

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 22:

CC TCC ACT CAG TGT GAT TCC AC 22

(24) INFORMATION FOR SEQ ID NO: 23:

20

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 35 bases

25

(B) TYPE: Nucleic acid

(C) STRANDEDNESS: Single

(D) TOPOLOGY: Linear

30

(ii) MOLECULE TYPE: Genomic DNA

(A) DESCRIPTION: p53, 35 bases flanking codon 175 of the gene for p53

35

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 23:

ATG AGG GAG GTT GTG AGG CGC TGC CCC CAC CAT GA 35

40

(25) INFORMATION FOR SEQ ID NO: 24:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 16 bases

45

(B) TYPE: Nucleic acid

(C) STRANDEDNESS: Single

50

(D) TOPOLOGY: Linear

5

(ii) MOLECULE TYPE: Genomic DNA

(A) DESCRIPTION: p53.5', The sequence of 16 bases
extending 5' from the C of codon 175 of the p53 gene.

10

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 24:

TC ATG GTG GGG GCA GC 16

15

(26) INFORMATION FOR SEQ ID NO: 25:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 19 bases

20

(B) TYPE: Nucleic acid

(C) STRANDEDNESS: Single

25

(D) TOPOLOGY: Linear

(ii) MOLECULE TYPE: Genomic DNA

(A) DESCRIPTION: p53.3', The sequence of 19 bases
extending 3' from the G of codon 175 of the p53 gene.

30

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 25:

G CCT CAC AAC CTC CCT CAT 19

35

(27) INFORMATION FOR SEQ ID NO: 26

(i) SEQUENCE CHARACTERISTICS

40

(A) LENGTH: 45 bases

(B) TYPE: Nucleic acid

(C) STRANDEDNESS: Single

45

(ii) MOLECULE TYPE: Genomic DNA

(A) DESCRIPTION: R553X, bases 1763-1807 of exon 11 of the
CFTR gene with T substituted for C at base 1789

50

5 (ix) SEQUENCE DESCRIPTION SEQ ID NO 26

GT GGA ATC ACA CTG AGT GGA GGT CAA TGA GCA AGA ATT TCT TTA G 45

10 (28) INFORMATION FOR SEQ ID NO: 27

(i) SEQUENCE CHARACTERISTICS

(A) LENGTH: 24 bases

15 (B) TYPE: Nucleic acid

(C) STANDEDNESS: Single

20 (ii) MOLECULE TYPE: Genomic DNA

(A) DESCRIPTION: CF1, bases 1656-1678 of exon 10 of the CFTR

25 (ix) SEQUENCE DESCRIPTION SEQ ID NO 27

CT ATA TTC ATC ATA GGA AAC ACC A 24

What is claimed is:

1. A method for identifying a target polynucleic acid sequence comprising:
 - 5 a. selecting 2 probes, such that
 1. when the 2 probes are ligated, they are complementary to some or all of the suspected sequence of said target polynucleic acid,
 - 10 2. one of the probes is joined to a moiety that permits this probe to be easily separated from the reaction mixture, and
 3. the other probe is joined to a label,
- b. mixing the probes with the target polynucleic acid
 - 15 in such a manner that the probes will hybridize to the target polynucleic acid,
 - c. adding a ligating reagent,
 - d. denaturing the reaction mixture so that the probe will be separated from the target polynucleic acid,
 - 20 e. separating the probe, utilizing the moiety that permits separation, and
 - f. analyzing the separated probe to determine if the label is attached thereto.
- 25 2. A method for identifying a target polynucleic acid sequence comprising:
 - a. selecting 2 probes, such that
 - 30 1. if the 2 probes were ligated, they would be complementary to some or all of the suspected sequence of said target polynucleic acid,
 2. one of the probes is joined to a moiety that permits this probe to be easily

- separated from the reaction mixture, and
3. the other probe is joined to a label,
- b. mixing the probes with the target polynucleic acid
in such a manner that the probes will
- 5 hybridize to the target polynucleic acid,
- c. separating the probe, utilizing the moiety that
permits separation, and
- d. analyzing the separated probe to determine if the
label is attached thereto.
- 10
3. A method of claim 1 wherein the moiety that
permits the probe to be separated is an insoluble
particle.
- 15
4. A method of claim 1 wherein the moiety that
permits the probe to be separated is a magnetic
particle.
- 20
5. A method of claim 1 wherein the label is selected
from the group consisting of enzymatic moieties,
radioactive moieties, fluorescent moieties,
luminescent moieties, and entities that permit
subsequent attachment to a label.
- 25
6. A method of claim 5 wherein said entities that
permit subsequent attachment to a label are avidin
or biotin.
- 30
7. A method of claim 5 wherein the label is a
luminescent material.
8. A method of claim 7 wherein the label is an
acridinium ester.

- 5 9. A method of claim 1 in which the label is an acridinium ester and the analysis of the separated probe to determine the presence of the label comprises the addition of DNAase before addition of the flash reagent.
- 10 10. A method of claim 1 wherein the target polynucleic acid is selected from the group consisting of DNA or polymers thereof, RNA or polymers thereof, and viral material.
- 15 11. A method of claim 1 which also includes a step for amplifying the target polynucleic acid before it is mixed with said probes.
- 20 12. A method of claim 11 in which the amplification technique is selected from the group consisting of polymerase chain reaction, ligase chain reaction and QB replicase.
- 25 13. A method of claim 1 in which the ligating agent acts enzymatically or chemically to join the two probes.
- 30 14. A method of claim 13 in which the ligating agent is ligase.
15. A method of claim 1 in which sodium chloride is present during the ligating step, such sodium chloride being present at a concentration of 200 - 1000 mM.
16. A method of claim 15 in which the sodium chloride concentration is 500 - 700 mM.

17. A method of claim 16 in which the sodium chloride concentration is approximately 600 mM.
18. A method of claim 1 in which, after denaturation,
5 the sample is passed through a chromatography column, said column then being analyzed to determine if the label is attached thereto.
19. A method for identifying a suspected mutation in a
10 target DNA comprising:
a. selecting 2 probes, such that
1. if the 2 probes were ligated, they would be
complementary to some or all of the
suspected sequence of said target DNA,
15 2. one of the probes is joined to a moiety that
permits this probe to be easily
separated from the reaction mixture, and
3. the other probe is joined to a label,
b. mixing the probes with the target DNA in such a
20 manner that the probes will hybridize to the
target polynucleic acid,
c. separating the probe, utilizing the moiety that
permits separation, and
d. analyzing the separated probe to determine if the
25 label is attached thereto.
20. A method for identifying a suspected mutation in a
target DNA comprising:
a. selecting 2 probes, such that
30 1. when the 2 probes are ligated, they are
complementary to some or all of the
suspected sequence of said target DNA,
2. one of the probes is joined to a moiety that
permits this probe to be easily

- separated from the reaction mixture, and
3. the other probe is joined to a label,
 - b. mixing the probes with the target DNA in such a manner that the probes will hybridize to the target polynucleic acid,
 - c. adding a ligating reagent,
 - d. denaturing the reaction mixture so that the probe will be separated from the target DNA,
 - e. separating the probe, utilizing the moiety that permits separation, and
 - f. analyzing the separated probe to determine if the label is attached thereto.
21. A method of claim 20 wherein the mutation is one of several that could occur in the target DNA.
 22. A method of claim 21 wherein several different probes are used, each being specific to one of the expected target sequences and each using a label or moiety that permits separation such that the analysis of the separated probes permits identification of the sequence or sequences present in the target.
 23. A method for identifying a target polynucleic acid sequence comprising:
 - a. selecting 2 probes, such that
 1. when the 2 probes are ligated, they are complementary to some or all of the suspected sequence of said target polynucleic acid,
 2. one of the probes is joined to a subunit of the midivariant sequence, and
 3. the other probe is joined to the second

- subunit of midivariant,
- b. mixing the probes with the target polynucleic acid in such a manner that the probes will hybridize to the target polynucleic acid,
 - 5 c. adding a ligating reagent,
 - d. optionally denaturing the reaction mixture so that the probe will be separated from the target polynucleic acid,
 - 10 e. analyzing the separated probe to determine if said probe had the ability to replicate if put in a reaction mixture containing QB replicase.
24. The method of claim 23 in which QB replicase is added along with the probes, and the analysis comprises determining if the probes had replicated.
- 15 25. A method for identifying whether the sequence of a target polynucleic acid is complementary to that of two or more probes comprising:
- 20 a. selecting said probes, such that
 - 1. when the probes are ligated, they are complementary to some or all of the suspected sequence of said target polynucleic acid,
 - 25 2. one of the terminal probes is joined to a moiety that permits this probe to be easily separated from the reaction mixture, and
 - 30 3. the other terminal probe is joined to a label,
 - b. mixing the probes with the target polynucleic acid in such a manner that the probes will hybridize to the target polynucleic acid,

- c. adding a ligating reagent,
 - d. denaturing the reaction mixture so that the probe will be separated from the target polynucleic acid,
 - 5 e. separating the probe, utilizing the moiety that permits separation, and
 - f. analyzing the separated probe to determine if the label is attached thereto.
- 10 26. A method of claim 1 in which the label is located in such a position that it does not interfere with hybridization and ligation.
- 15 27. A method of claim 1 which also includes the process wherein, before denaturation, an aliquot is removed, said aliquot being analyzed to determine if the label is hybridized to the target by
- 20 a. separating the probe containing the moiety that permits separation and the other entities attached thereto and
 - b. analyzing said separated probe to determine if the label is attached thereto.
- 25 28. A method of claim 1 which also includes analyzing the supernatant remaining after separation of said probe containing said moiety that permits separation to analyze said supernatant for the presence of the label contained therein.
- 30 29. A method of claim 1, 14, 20 or 23 wherein the ligating reagent is selected from the group consisting of Taq DNA-ligase or T4 DNA-ligase.

30. A method of claim 30 in which the buffer containing said Taq DNA ligase also includes therein tRNA.
- 5 31. A method of claim 1, 20, 23 or 25 wherein the ligating reagent is added at the same time as the probes.

ATATAAGTAGTATCCTTTGTGGTTACTATAAAAGAAATTACCAAGGT
 PMP-SPACER-TATATTCATCATAGGAAACACCAATGATATTTCTTTAATGGTGCCA
 DMAE

ATATAAGTAGTATCCTTTGTGGTTTCTACTATAAAAGAAATTACCACGGT
 ::
 PMP-SPACER-TATATTCATCATAGGAAACACCA ATGATATTTTCTTTAATGGTGCCA
 DMAE
 508.CF-DMAE

ATATAAGTAGTATCCTTTGTGGTTACTATAAAAGAAATTACCACGGT
 :::
 PMP-SPACER-TATATTCATCATAGGAAACACCAATGATATTTTCTTTAATGGTGCCA
 A
 A
 G

508.NOR-LEAE LEAE

ATATAAGTAGTATCCTTTGTGGTTTCTACTATAAAAGAAATTACACGGT
 :::
 PMP-SPACER-TATATTCATCATAGGAAACACCAAAGATGATATTTCTTTAATGGTGCCA
 508.NOR-LEAE
 AE

1/20

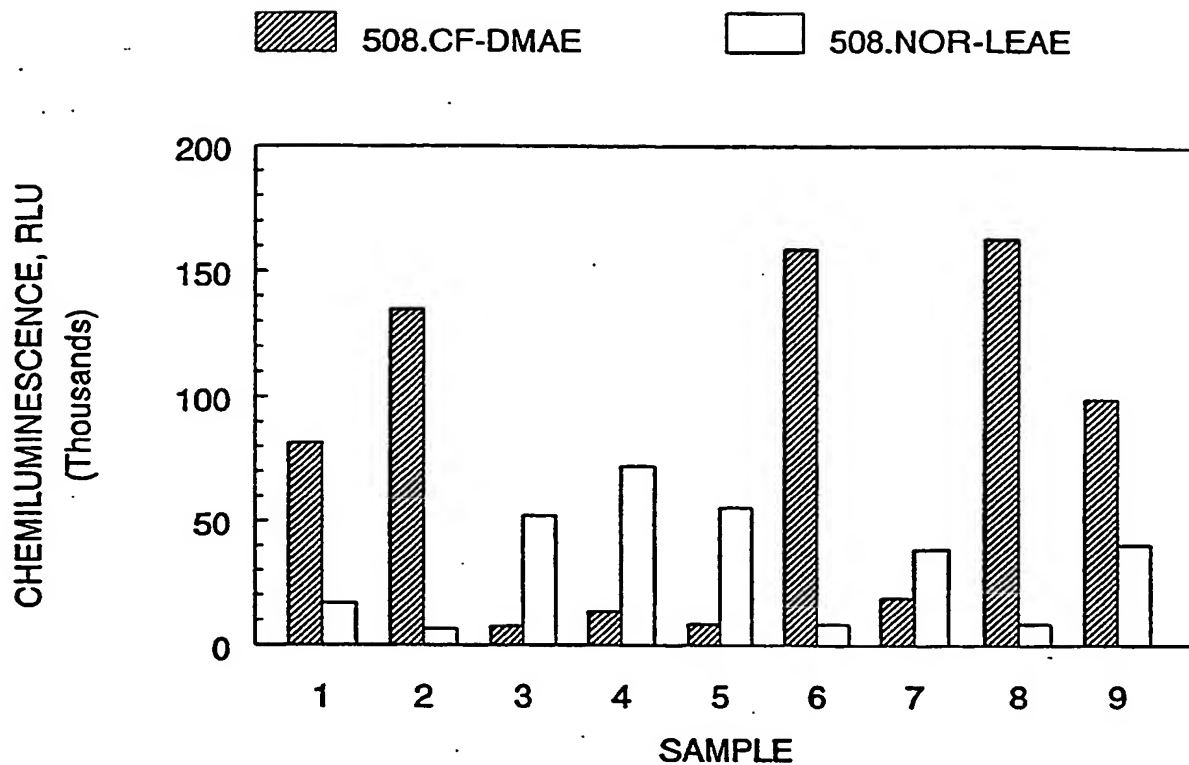


Figure 2

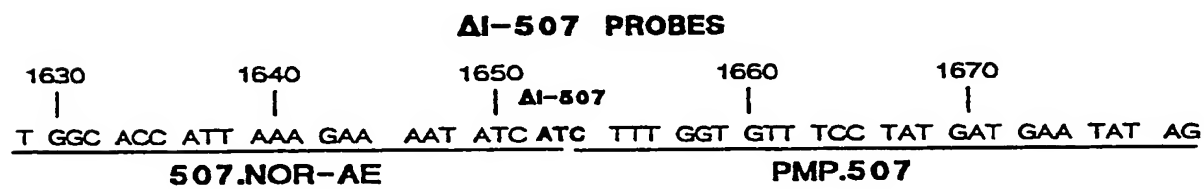
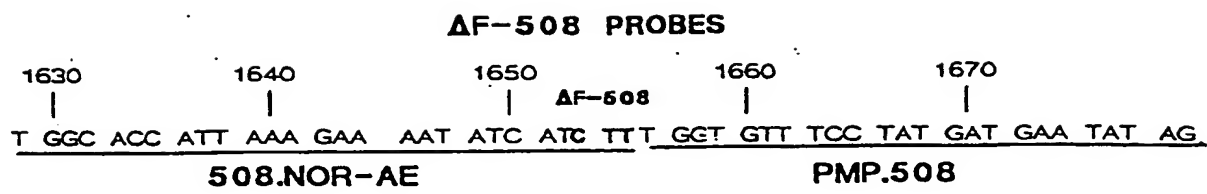


Figure 3

HYBRIDIZATION OF $\Delta F-508$ PROBES				HYBRIDIZATION OF $\Delta I-507$ PROBES			
NORMAL				NORMAL			
5' PMP	TGGTTTCTACTATAA ACCA <u>ATGATATT</u>	3' AE		5' PMP	TGGTTTCTACTATAA ACCAAAG <u>ATATT</u>	3' AE	
$\Delta F-508$				$\Delta F-508$			
5' PMP	TTGTGGTTACTATAA AACACCAAT <u>GATATT</u>	3' AE		5' PMP	TTGTGGTTACTATAA AACACCAAAG <u>GATATT</u>	3' AE	
$\Delta I-507$				$\Delta I-507$			
5' PMP	TGGTTTCTATAAAAG ACCAAT <u>GATATTTTC</u>	3' AE		5' PMP	TGGTTTCTATAAAAG ACCAAAGATATTTTC	3' AE	

Figure 4

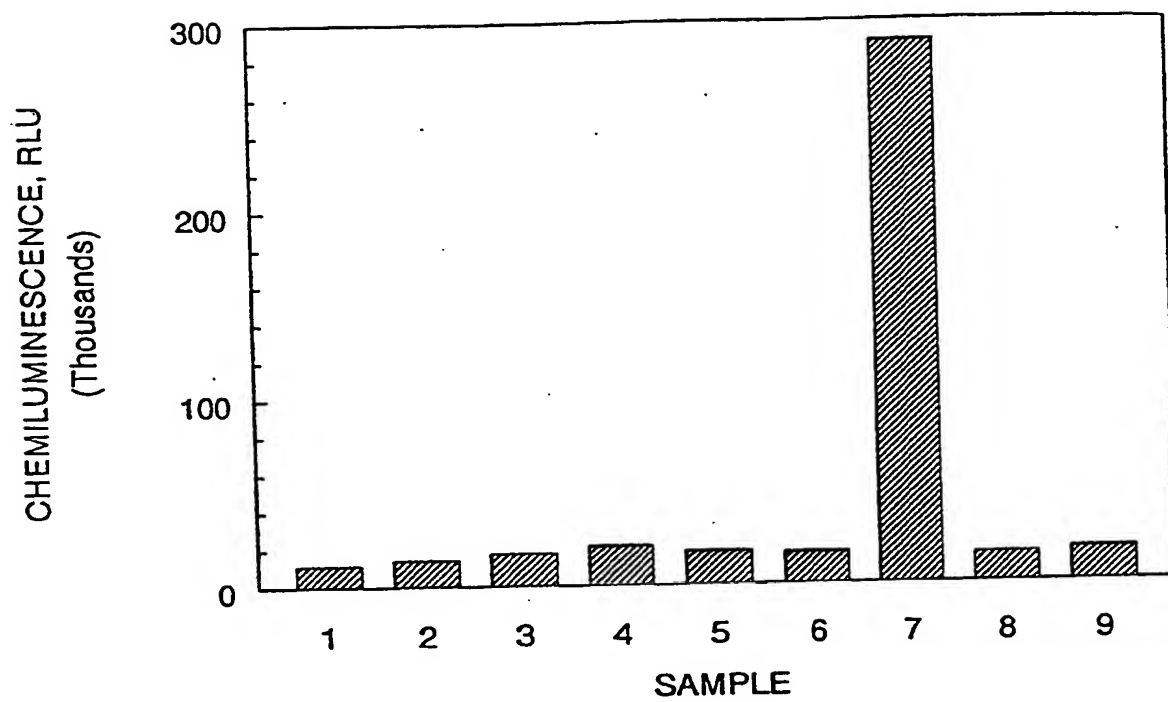


Figure 5

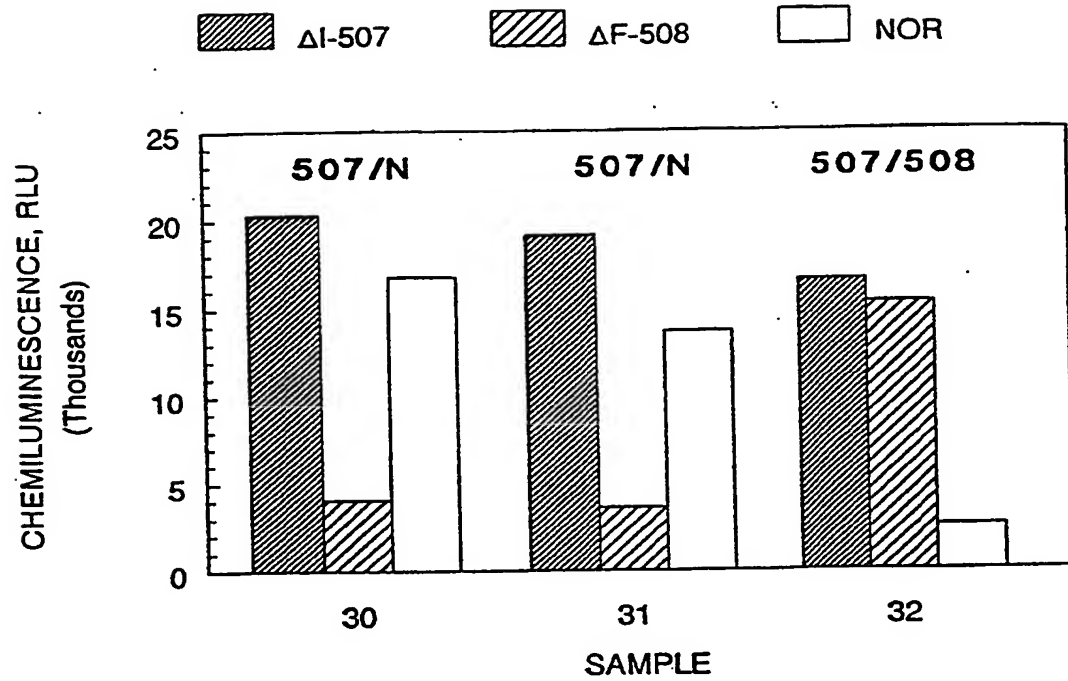


Figure 6

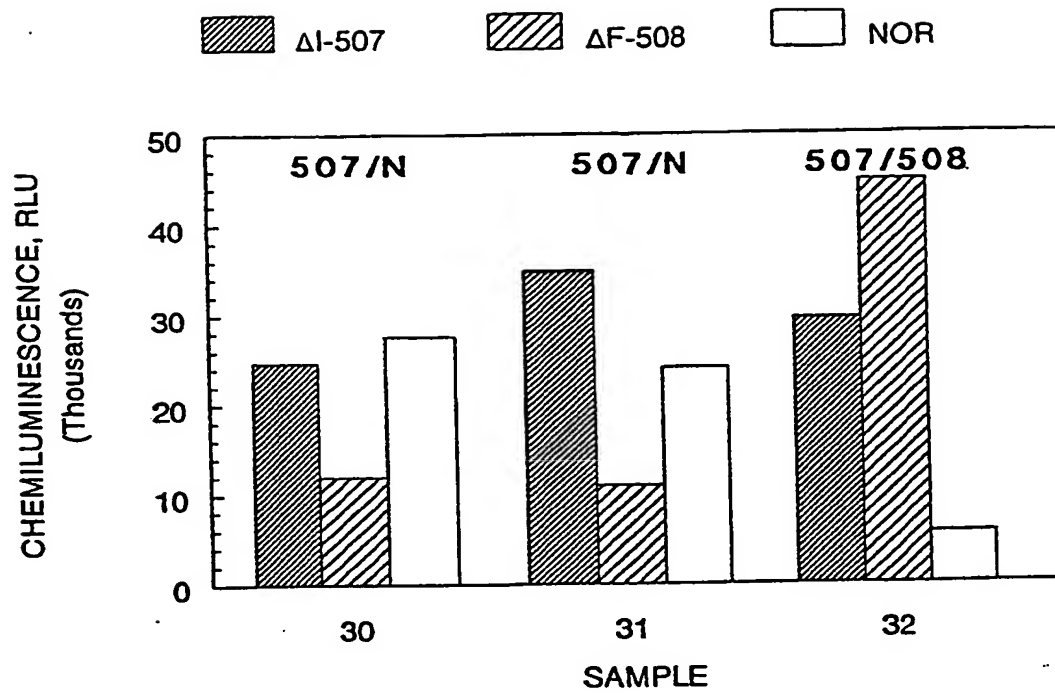
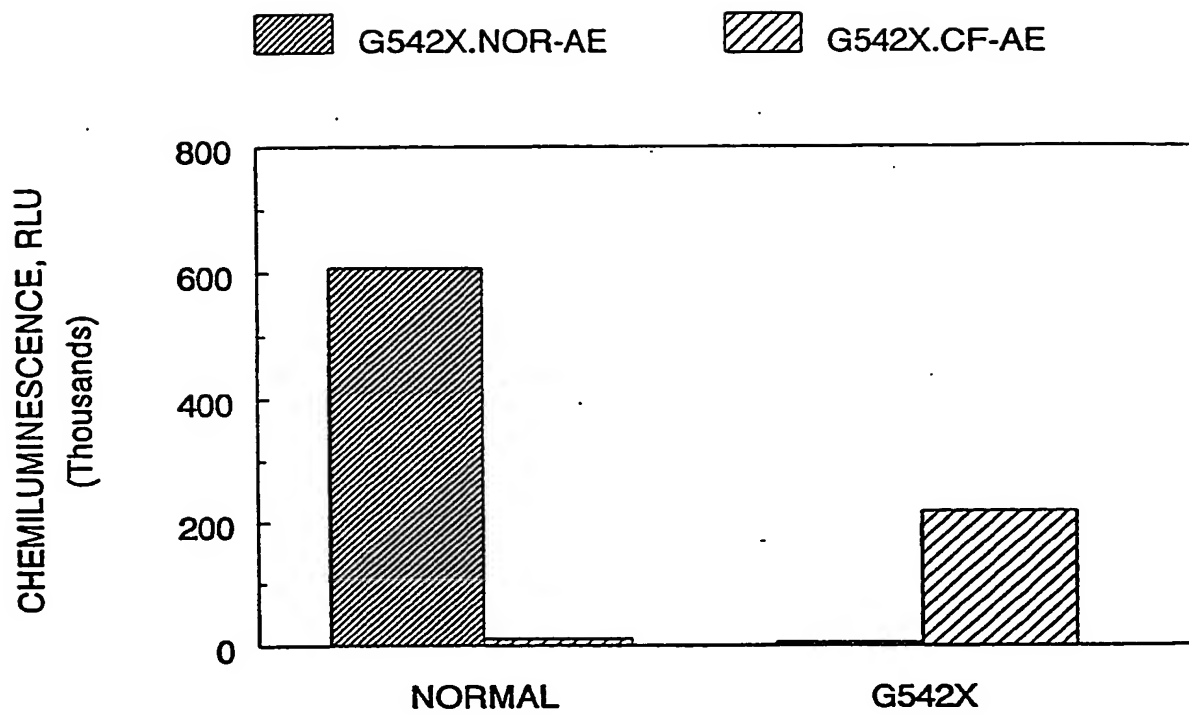


Figure 7



TARGET

Figure 8

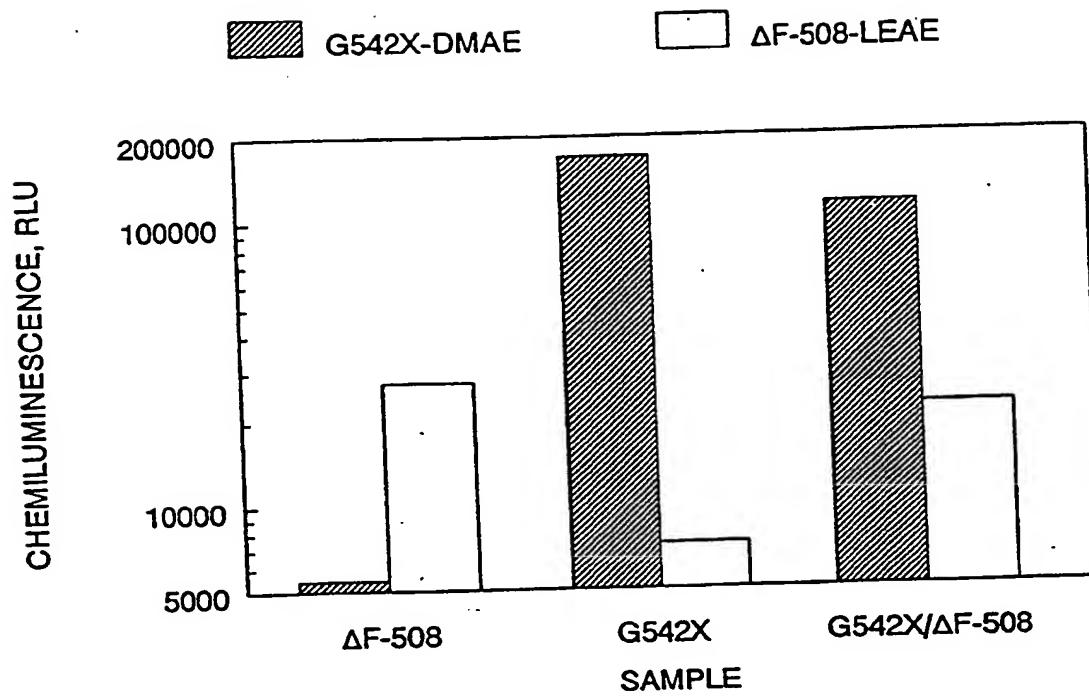


Figure 9

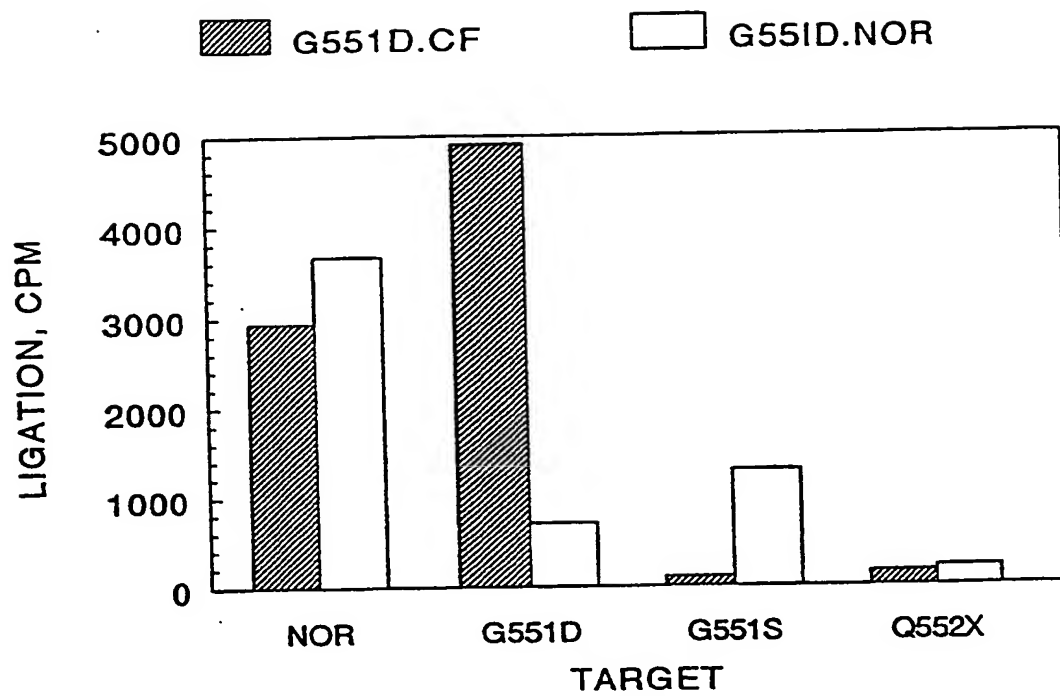


Figure 10

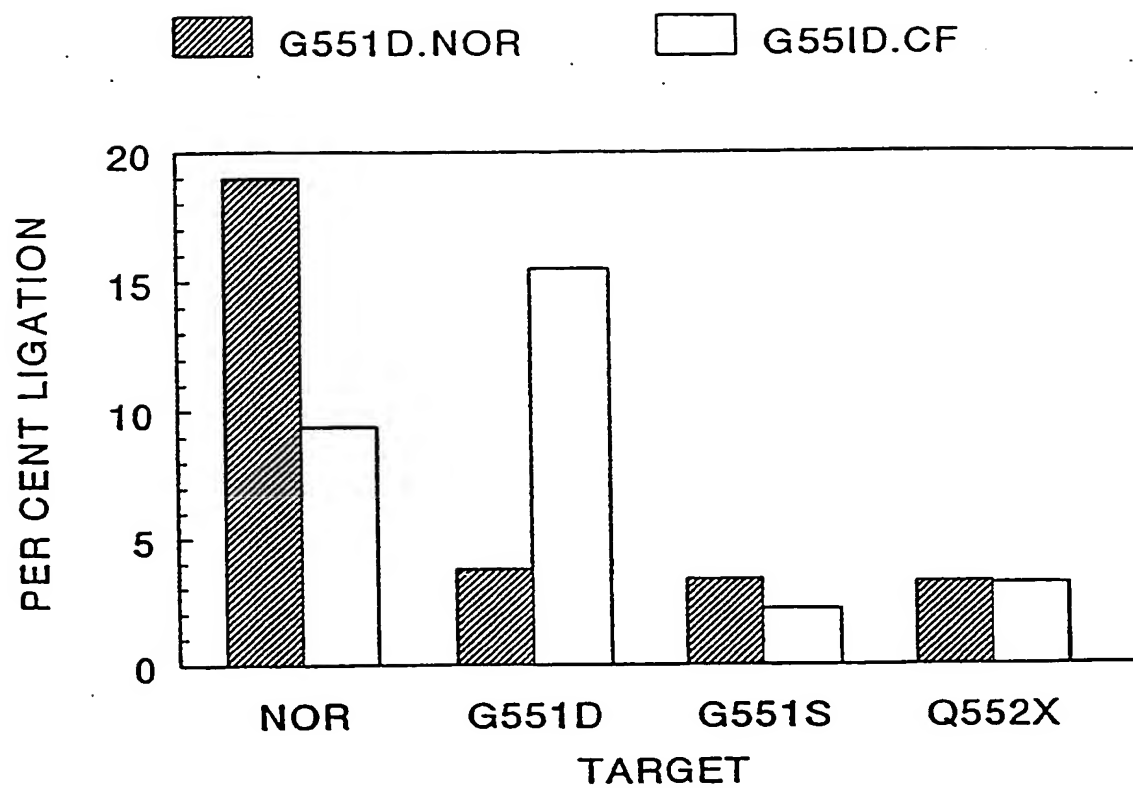
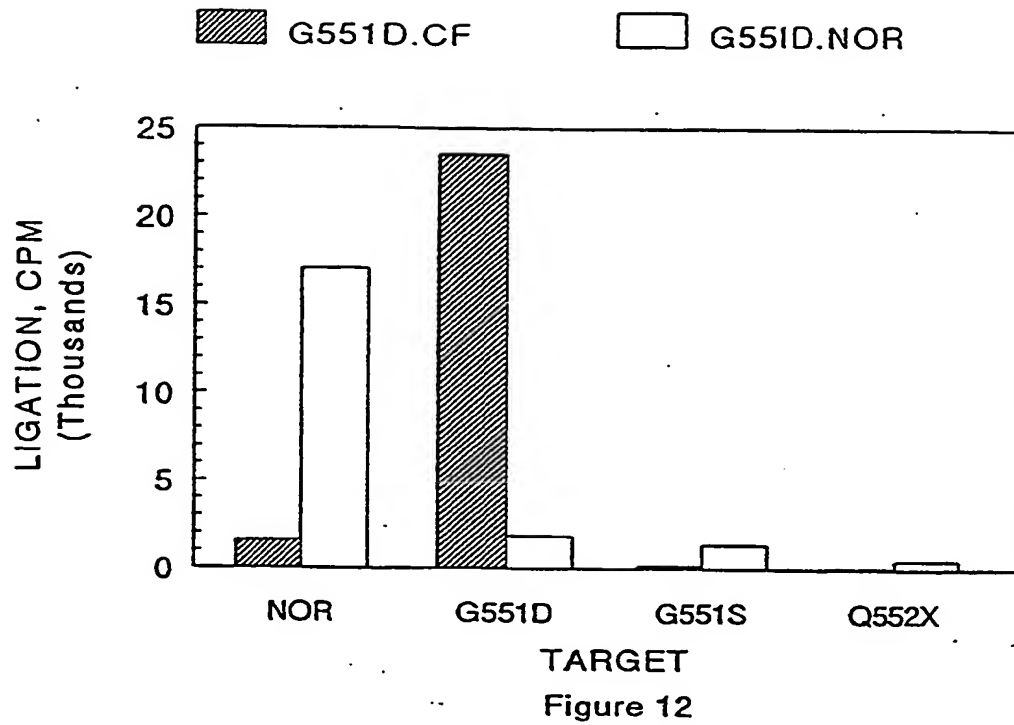


Figure 11



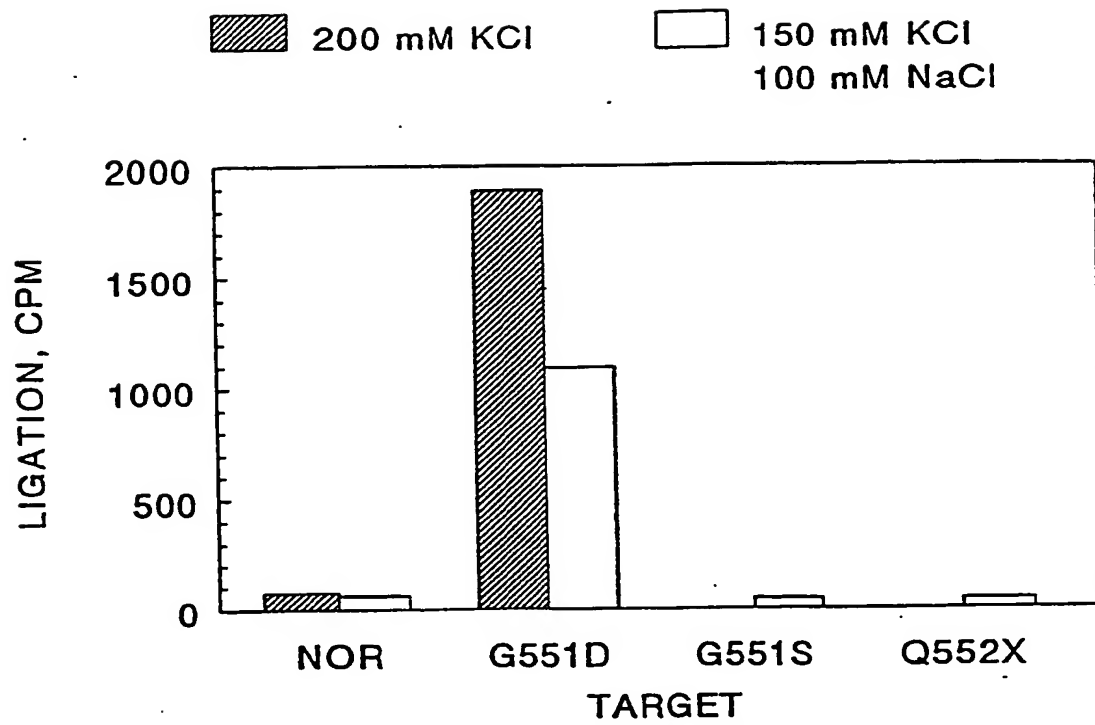


Figure 13

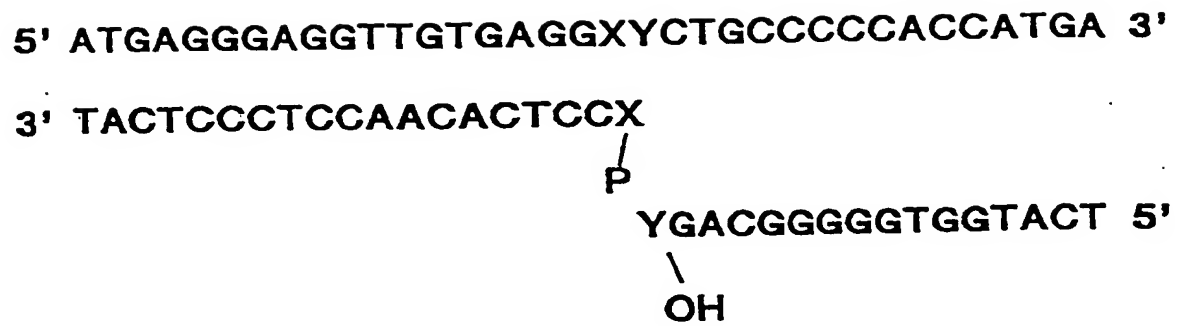


Figure 14

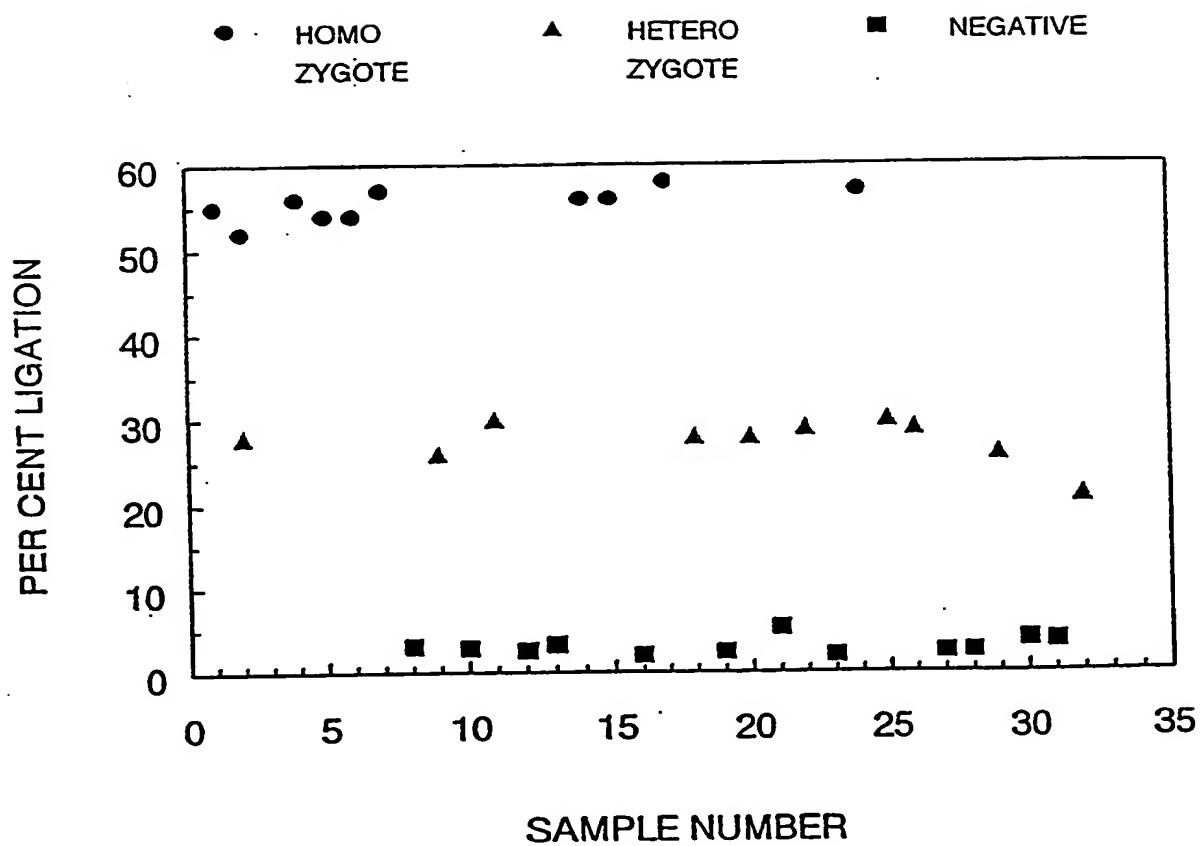
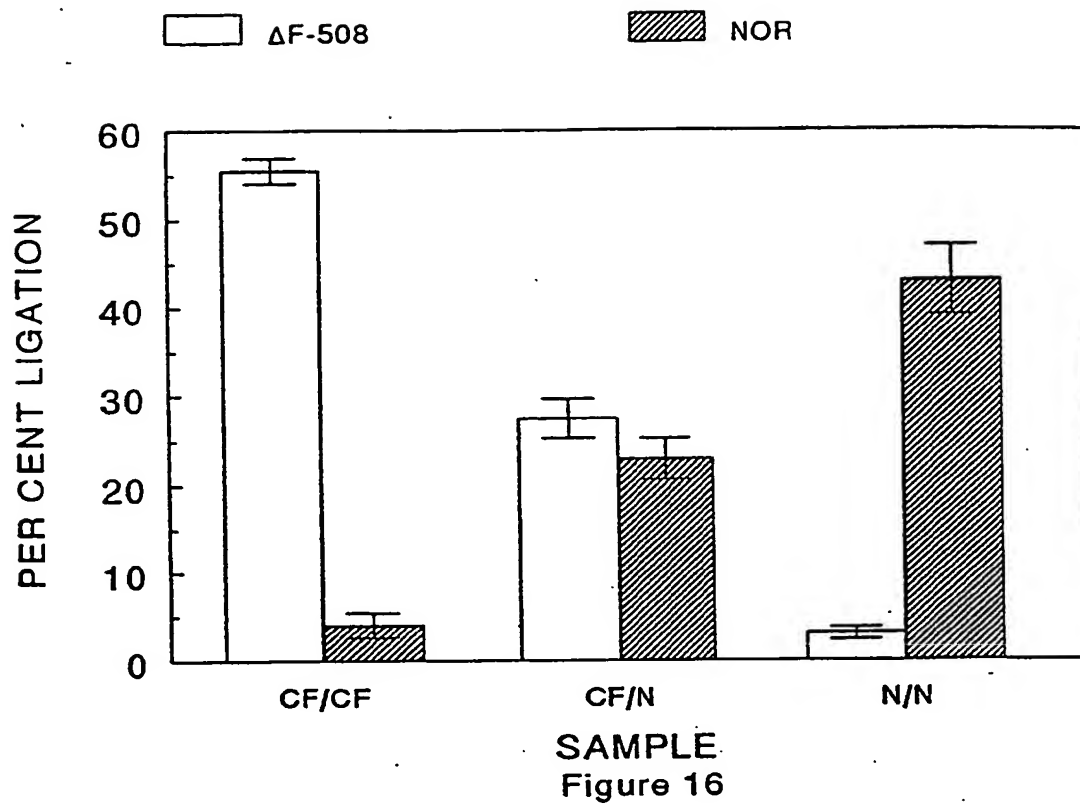
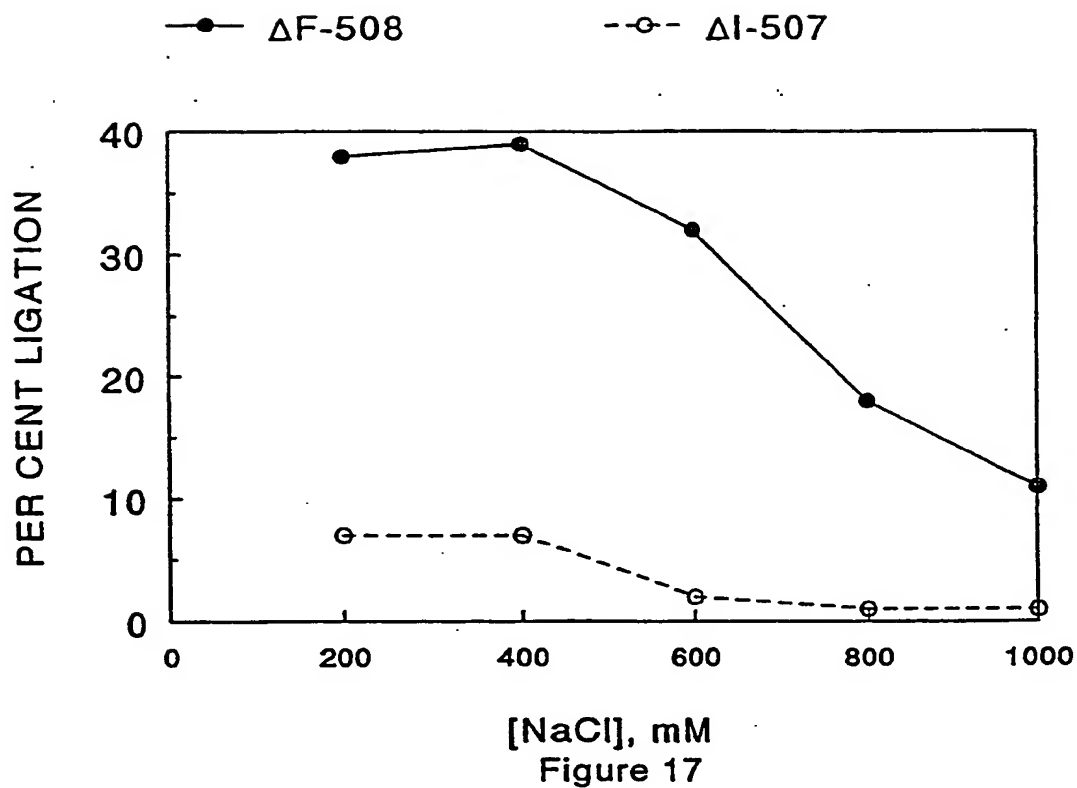


Figure 15





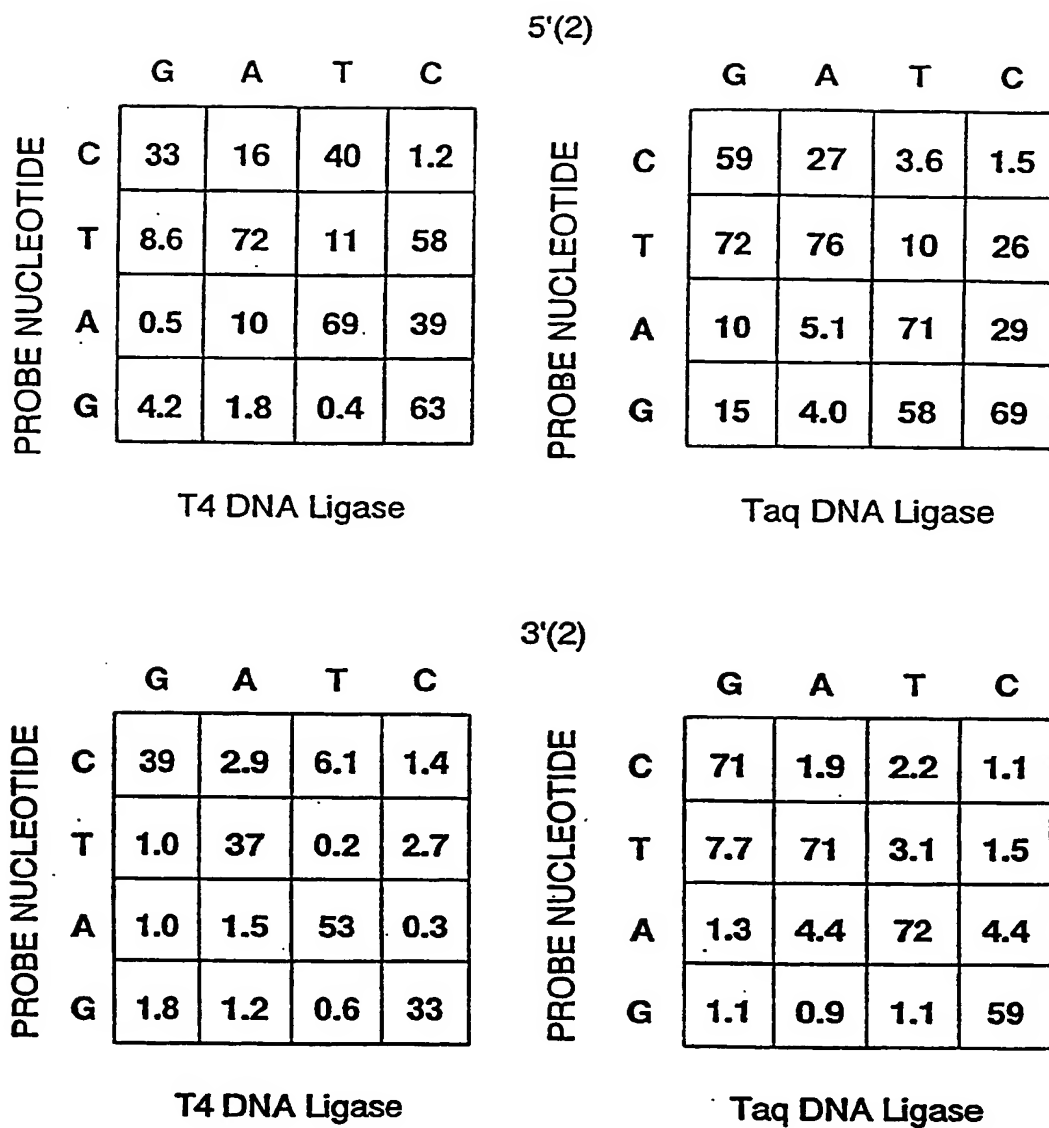


Figure 18

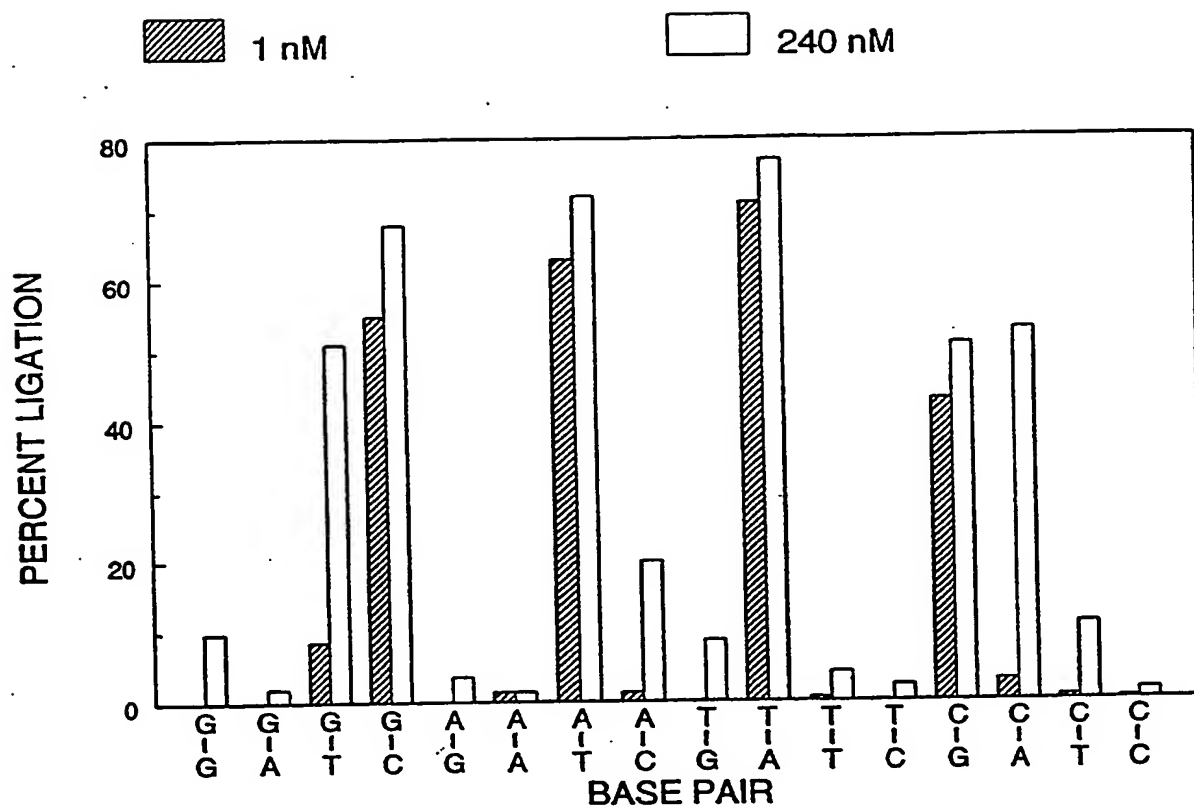


Figure 19

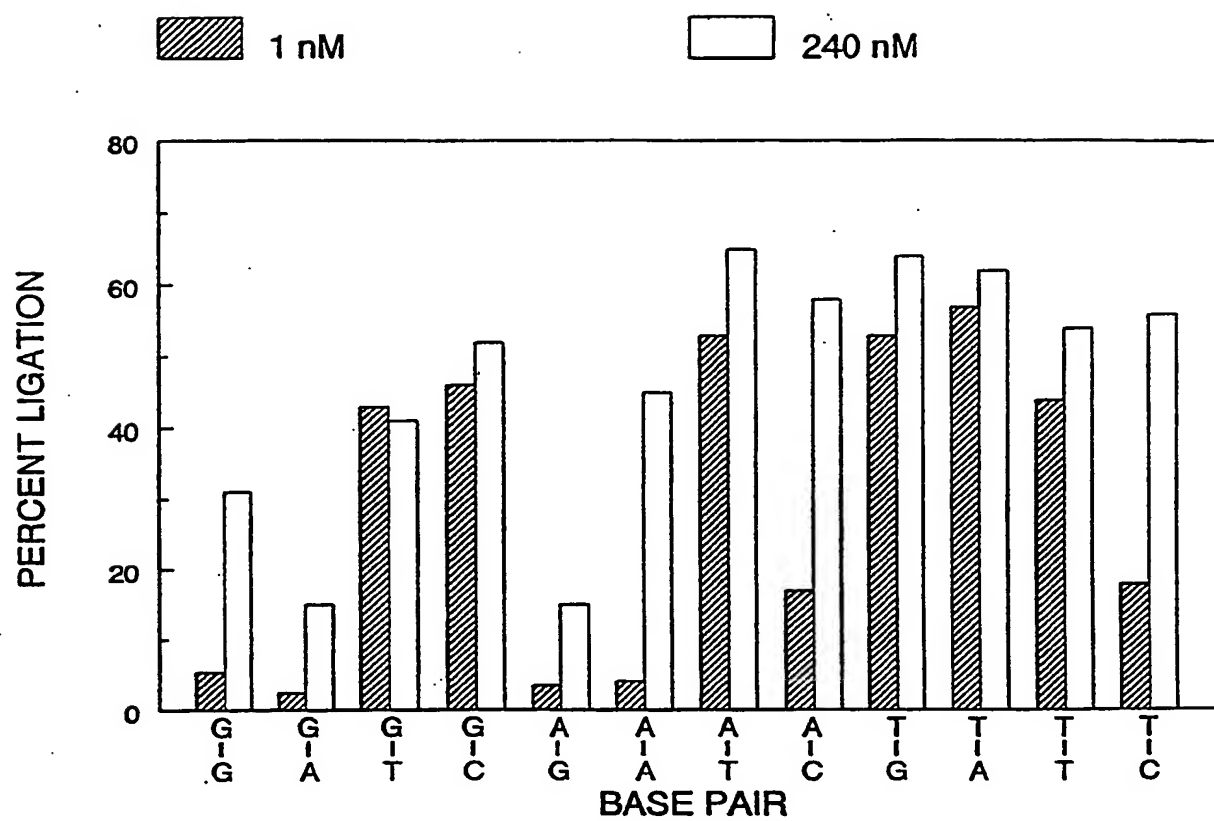


Figure 20

INTERNATIONAL SEARCH REPORT

International Application No

PCT/IB 95/00226

A. CLASSIFICATION OF SUBJECT MATTER
 IPC 6 C12Q1/68 C12P19/34 C07H21/04

According to International Patent Classification (IPC) or to both national classification and IPC

B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)
 IPC 6 C12Q

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

Electronic data base consulted during the international search (name of data base and, where practical, search terms used)

C. DOCUMENTS CONSIDERED TO BE RELEVANT

Category *	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
X	EP,A,0 185 494 (APPLIED BIOSYSTEMS INC.) 25 June 1986	1-10, 13, 14, 18-21
Y	see the whole document	12, 15-17, 22-31
Y	EP,A,0 481 704 (CIBA CORNING DIAGNOSTICS CORP) 22 April 1992 see claims 1-15; figures 2,3	1-31
Y	WO,A,93 20239 (APPLIED BIOSYSTEMS) 14 October 1993 see claims 1-12	1-31

☒ Further documents are listed in the continuation of box C.

☒ Patent family members are listed in annex.

* Special categories of cited documents:

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Date of the actual completion of the international search

24 August 1995

Date of mailing of the international search report

19.09.95

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Osborne, H

INTERNATIONAL SEARCH REPORT

International Application No

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C.(Continuation) DOCUMENTS CONSIDERED TO BE RELEVANT

Category *	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
Y	PROC. NATL. ACAD. SCI US, vol. 87, October 1990 pages 8923-27, NICKERSON, D. ET AL. 'automated DNA diagnostics using an elisa-based oligonucleotide assay' see the whole document ----	1-31
Y	EP,A,0 552 545 (PIONEER HI-BREED) 28 July 1993 see the whole document ----	1-31
Y	EP,A,0 336 731 (CITY OF HOPE) 11 October 1989 see claims 11,29 ----	15-17
T	WO,A,94 08047 (ABBOTT LABS.) 14 April 1994 see page 3, line 11 - page 4, line 12 -----	15-17

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Information : Patent family members

International Applicant No

PCT/IB 95/00226

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		DE-T- 68915202	29-09-94
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